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RESEARCH ARTICLE

IDENTIFICATION OF SOMATIC EMBRYOGENESIS RECEPTOR KINASE (SERK) GENES IN SORGHUM THROUGH IN SILICO ANALYSIS

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ABSTRACT

Somatic embryogenesis receptor kinases (SERKs) constitute a small gene family and are functionally conserved in plants with a specific role in embryogenesis and possibly in other developmental processes. The present investigation was aimed at identifying the SERK gene (s) present in sorghum genome through in silico studies. Here we report two SERK genes (SbSERK1, and SbSERK2) from sorghum (Sorghum bicolor (L.) Moench.) by the comparative analysis of known SERK cDNA sequences from sorghum and other plants and their chromosomal location on sorghum. The sequences of SbSERK1 and SbSERK2 were more similar to that of ZmSERK1 and ZmSERK2 genes of maize. A putative SbSERK3 was also identified which was more similar to the rice OsSERK1.

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INTRODUCTION

Sorghum (Sorghum bicolor (L.) Moench.) is a leading cereal, ranking fifth in importance after wheat, rice, maize, and barley. It is a self-pollinated diploid (2n=2x=20) C4 grass widely adapted to diverse agricultural environments around the world. As sorghum has a smaller genome size of 730 Mbp and is fully sequenced, it makes an attractive model for functional genomics of C4 grasses (Paterson et al., 2009). Somatic Embryogenesis (SE) is an in vitro asexual reproduction process in which somatic cells gives raise to somatic embryos under favorable experimental conditions (Madhu et al., 2015). In tissue culture, differentiated somatic cells acquire embryogenic competence and proliferate as embryogenic cells during the induction phase (Dodeman et al., 1997 and Parameswari Namasivayam., 2007). The events that take place during the period in which plant cells undergo the transition from somatic to embryogenic cell are poorly understood (De Jong et al., 1993). Genes like LEC1 (Lotan et al., 1998), LEC2 (Stone et al., 2001), BBM (Boutilier et al., 2002), WU (Zuo et al., 2002) and AGL15 (Harding et al., 2003) are involved in the somatic embryogenesis process but

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in the later stages of SE. These genes inturn helps in increasing the overall frequency of SE and prolongs the process of SE. However out of all the genes that have been isolated and analyzed during SE, somatic embryogenesis receptor kinase (SERK) gene, is the only one that has been shown successfully to be a specific marker distinguishing individual embryo forming cells in carrot suspension cultures (Schmidt *et al.*, 1997). SERKs constitute a small gene family and are functionally conserved in plants with a specific role in embryogenesis and possibly other developmental processes. In addition to play as a key role in embryogenesis process, SERK gene is also involved in the transduction of extracellular signaling processes as diverse as plant development, disease resistance or self incompatibility (Baudino *et al.*, 2001; Krupa *et al.*, 2006).

SERK belongs to Leucine-rich repeat, receptor like kinases (LRR-RLKs), a subgroup of protein kinases. The predicted SERK protein contains an N-terminal Leucine zipper (LZ) domain followed by 5 LRRs, a serine and proline rich SPP domain, a transmembrane domain and an intracellular serine /threonine kinase domain (Figure 1). The SPP domain is a unique feature of the SERK family of receptor kinases (Schmidt et al., 1997; Hecht et al., 2001). A unique feature of SERK protein is the presence of proline-rich region between the extracellular LRR domain of SERK and the membrane-

spanning region, which is a conserved feature of plant cell wall proteins known as extensions (Varner and Lin 1989). Upto now, the ubiquitous presence of this small *SERK* gene family has been identified, isolated and characterized in almost all the plant species. So in this current study, based on the c-DNA sequence information obtained from the close relatives of sorghum like maize and rice, attempt was made to predict the loci, structure and copy number of *SERK* genes on sorghum genome using *in silico* studies.

MATERIALS AND METHODS

To predict the presence of *SERK* genes in sorghum, Phytozome *v* 10.3 is used as a basic platform which serves as the best plant comparative genomic portal. Nucleotide sequences were compared to EMBL and GenBank databases using the BLASTN (nucleotide query to nucleotide db) algorithm. Clustal Omega tool is used for the protein sequence analysis which inturn uses seeded guide trees and HMM profile-profile techniques to generate alignments between three or more protein sequences. NCBI database is used for identification of c-DNA and its relative protein sequence information of Maize, Rice, Arabidopsis and Medicago. Phylogenetic trees (Figure 2) were constructed using ClustalW of EMBL database.

RESULTS AND DISCUSSION

Based on the *SERK* gene c-DNA sequence information from other crop plants like maize and rice, genome wide comparative analysis was performed to identify the loci and copy number of *SERK* genes in sorghum. The main referral gene sequences used in this study are *ZmSERK1* (EMBL accession number AJ400869) and *OsSERK1* (EMBL accession number AB188247).

SERK gene family

SERK belongs to a small gene family with at least five members in Arabidopsis (Hecht et al., 2001), three in maize (Baudino et al., 2001), five in Medicago truncatula (Nolan et al., 2003), four in Helianthus annuus (Thomas et al., 2004), two in rice (Ito et al., 2005; Hu et al., 2005) and at least three in wheat (Singla et al., 2008). A correlation between SERK expression and somatic embryogenesis was demonstrated in cultured tissues of carrot (Schmidt et al., 1997), Dactylis glomerata (Somleva et al., 2000), Arabidopsis thaliana (Hecht et al., 2001), Medicago truncatula (Nolan et al., 2003), sunflower (Thomas et al., 2004), rice (Hu et al., 2005), cocoa (Santos et al., 2005), and Triticum aestivum (Singla et al., 2008).

The SbSERK family

The SERK gene family in sorghum has at least three members. These three genes were identified during a screen of SERK c-DNA sequences of maize and rice with sorghum. ZmSERK 1 and 2 helped in identifying two SERK genes in sorghum and one by OsSERK1. These three genes are precisely described here at their nucleotide and protein levels.

SbSERK1

SbSERK1 gene was identified while analyzing ZmSERK1 gene using BLASTN algorithm along the sorghum genome and hits a maximum similarity on chromosome (chr) 6 in sorghum. The total length of this functional gene is 5285 bp in which coding region occupies 1869 bp. The first portion of the gene starts with 284bp of 5'UTR and ends with 241 bp of 3'UTR and between these two UTR's it is occupied by 11 exons and 10 introns. When analyzed at the protein level using SIM module of ExPASy tools SbSERK1 having 622 amino acids (aa) is showing the highest similarity of 86.7% with the peptide sequence of ZmSERK1 gene containing 622 aa. The similarities of protein sequences between SbSERK1, ZmSERK1, OsSERK1, AtSERK1 and MtSERK1 are shown in Figure 3.



Figure 1. Structural features of SERK genes in sorghum

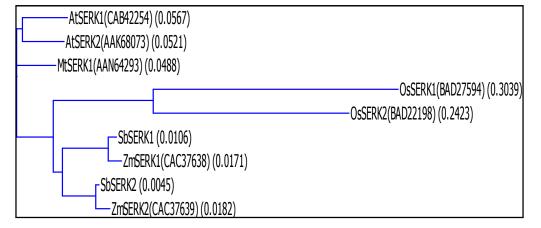


Figure 2. Phylogenetic relationship of SERK1 and SERK2 Proteins



Figure 3. Multiple sequence alignment of Sorghum SERK1

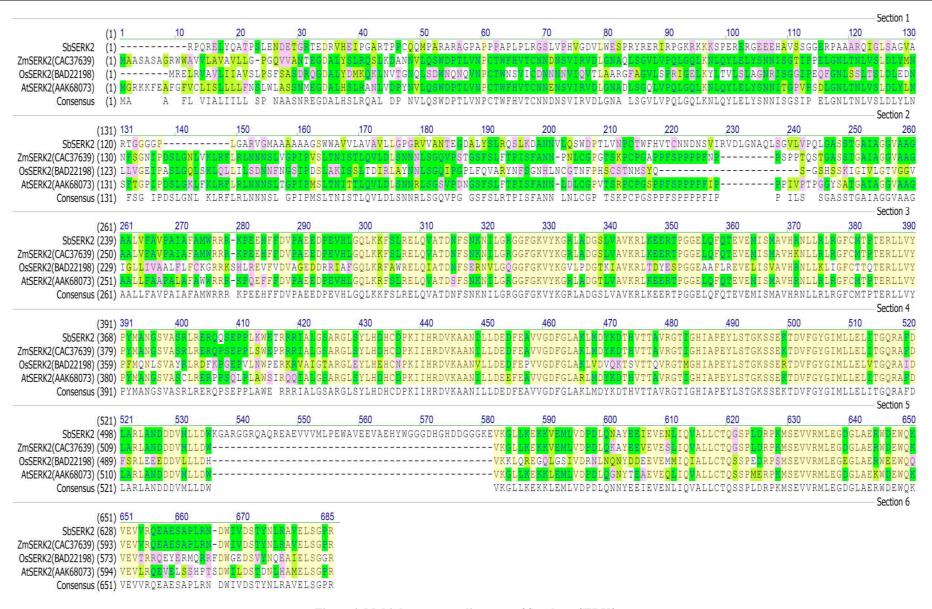


Figure 4. Multiple sequence alignment of Sorghum SERK2

SbSERK2

This specific *SbSERK2* gene was identified on *chr* 4 in sorghum genome which shows the highest similarity with *ZmSERK2* of maize. The total predicted length of this gene is 6152 bp having 353 bp of 5'UTR and 457 bp of 3'UTR between this occupied by a total of 11 exons and 10 introns. The CDS (coding sequence) sequence consists of 1881 bp which codes for 626 amino acids. The similarity of *SbSERK2* peptide sequence when analyzed with SIM module of ExPASy tools with the peptide sequence of *ZmSERK2* (626 aa) is 97.1%, which shows the close relative protein functions between sorghum and maize. The similarities of protein sequences between *SbSERK2*, *ZmSERK2*, *OsSERK2*, and *AtSERK1* are shown in Figure 4.

SbSERK3

SbSERK3 gene was identified while analyzing OsSERK1 gene using BLASTN algorithm along the sorghum genome and hits a maximum similarity on chr 7 in sorghum. The total length of this functional gene is 6597 bp in which coding region occupies 1896 bp. The first portion of the gene starts with 470 bp of 5'UTR and ends with 611 bp of 3'UTR and between these two UTR's it is occupied by 11 exons and 10 introns. When analyzed at the protein level using SIM module of ExPASy tools SbSERK1 having 631 amino acids (aa) is showing the highest similarity of 94.2% with the peptide sequence of OsSERK1 gene containing 620aa.

The available literature and expression studies still do not provide adequate information to differentiation between the several *SERK* genes in each species. The protein sequences among sorghum, maize and rice share sequence similarities up to 86.7-94.2% is well interpreted as essential for a common function. *ZmSERK1* is preferentially expressed in male and female reproductive tissues while *ZmSERK2* was relatively uniform in expression in the tissues investigated (Baudino *et al.*, 2001). The suppression of *OsSERK1* by RNA interference resulted in an inhibition and *SERK1* over expression resulted in induction of shoot regeneration from callus. Over expression of *OsSERK1* resulted in an increased resistance to blast fungus (Hu *et al.*, 2005).

In Arabidopsis, at ovule maturity, all cells of the embryo sac express AtSERK1, the SERK gene of Arabidposis (Hecht et al., 2001). The AtSERK1 gene is expressed megasporogenesis and in all cells of the embryo sac up to the stage of fertilization. After fertilization, AtSERK1 promoterdriven GUS activity is found in all cells of the developing embryo up to the heart stage. The expression of the AtSERK1 homolog gene *PpSERK1* was studied by Albertini *et al.* (2005) who revealed by in Situ hybridization that PpSERK is expressed in the megaspore mother cell of sexual genotypes, but not in that of apomictically reproducing Poa pratensis plants. Partial cDNA fragments showing homology to SERK and LRR-Kinase genes of Arabidopsis have also been isolated in buffel grass by subtractive hybridization (Dwivedi et al., 2005). It had very low abundance (100 times less) in apomicts while it is over-expressed in its close sexual relative (F2 segregant). Over expression of SERK1 in Arabidopsis did not result in any obvious plant phenotypes, but gave a 3 to 4-fold increase in embryogenic competence, which indicates that SERK1 not only marks embryogenic competence, but also promotes the transition of somatic cells to an embryogenic state (Hecht *et al.*, 2001). The identification of SERK genes in sorghum is expected to throw more light on its function in somatic embryogenesis, embryo development and other phenomena. The role of SERK genes in megasporogenesis (Albertini *et al.*, 2005; Dwivedi *et al.*, 2005) and microsporogenesis (Hecht *et al.*, 2001) may be further investigated to utilize them in development of apomictic and hybrid cultivars.

SP- Signal peptide domain; ZIP- N-terminal Leucine zipper domain; LRR- leucine rich repeat-Receptor Like kinases; SPP- serine and proline rich domain; TM- transmembrane domain; Kinase- an intracellular serine / threonine kinase domain; C- C terminal domain. Proline-rich SPP domain between LRR and TM is a unique feature of the SERK family of receptor kinases. The phylogenetic tree with the SERK family in sorghum (Sb), Arabidopsis (At), alfalfa (Mt), maize (Zm) and rice (Os) generated using ClustalW using amino acid sequence data.

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REFERENCES

Albertini, E., Marconi, G., Reale, L., Barcaccia, G., Porceddu, A., Ferranti, F. and Falcinelli, M. 2005. SERK and APOSTART. Candidate Gene for Apomixis in Poa pratensis. Plant Physiol., 138: 2185-2199.

Baudino, S., Hansen, S., Brettschneider, R., Hecht Valérie,
FG., Dresselhaus, T., Lorz, H., Dumas, C. and Rogowsky,
P.M. 2001. Molecular characterisation of two novel maize
LRR receptor-like kinases, which belong to the *SERK* gene family. *Planta*, 213:1-10.

Boutilier, K., Offringa, R., Sharma, V.K., Kieft, H., Ouellet, T., Zhang, L., Hattori, J., Liu, C.M., van Lammeren, A.A.M., Miki, B.L.A., Custers, J.B.M. and van Lookeren Campage, M.M. 2002. Ectopic expression of *BABY BOOM* triggers a conversion from vegetative to embryonic growth. *Plant Cell*, 14: 1737–1749

De Jong, A.J., Schmidt, E.D. and de Vries, SC. 1993. Early events in higher-plant embryogenesis. *Plant Mol. Biol.*, 22:367–377

Dodeman, V.L., Ducreux, G. and Kreis, M. 1997. Zygotic embryogenesis *versus* somatic embryogenesis. *J. Exp. Bot.*, 48: 1493–1509.

Dwivedi, K.K. 2005. Isolation and cloning of genes associated with apomixis in *Cenchrus ciliaris* L. Ph.D. Dissertation, Bundelkhand University, Jhansi. U.P.

Harding, E.W., Tang, W.N., Nichols, K.W., Fernandez, D.E. and Perry, S.E. 2003. Expression and maintenance of embryogenic potential is enhanced through constitutive expression of *AGAMOUS*-Like15. *Plant Physiol.*, 133, 653–663

- Hecht, V., Vielle-Calzada, J.P., Hartog, M.V., Schmidt, D.L.,
 Boutilier, K., Grossniklaus, U. and de Vries, S.C. 2001.
 The *Arabidopsis* somatic embryogenesis receptor kinase 1 gene is expressed in developing ovules and embryos and enhances embryogenic competence in culture. *Plant Physiol.*, 127: 803-816
- Hu, H., Xiong, L. and Ynag, Y. 2005. Rice *SERK1* gene positively regulates somatic embryogenesis of cultured cell and host defense response against fungal infection. *Planta*, 222:107-117.
- Ito, Y., Takaya, K. and Kurata, N.2005. Expression of *SERK* family receptor-like protein kinase genes in rice. *Biochim Biophys. Acta*, 1730:253-258
- Krupa, A., Anamika and Srinivasan, N. 2006. Genome-wide comparative analyses of domain organisation of repertoires of protein kinases of *Arabidopsis thaliana* and *Oryza sativa*. *Gene* 380:1–13
- Lotan, T., Ohto, M., Yee, K.M., West, M.A., Lo, R., Kwong, R.W., Yamagishi, K., Fischer, R.L., Goldberg, R.B. and Harada, J.J. 1998. Arabidopsis *LEAFYCOTYLEDON1* is sufficient to induce embryo development in vegetative cells. *Cell*, 93:1195–1205
- Madhu, P., Pushpa, K., Bhat B.V. and Balakrishna, D. 2015. Effect of kanamycin on explant selection in genetic transformation experiments of sorghum. *Intl. J. Sci. Res.*, 4(7): 30-33.
- Namasivayam, P. 2007. Acquisition of embryogenic competence during somatic embryogenesis. *Plant Cell Tissue Organ Cult.*, 90(1): 1–8.
- Nolan, K.E., Irwanto, R.R. and Rose, R.J. 2003. Auxin upregulates *MtSERK1* expression in both *Medicago truncatula* root forming and embryogenic cultures. *Plant Physiol.*, 133:218-230.
- Paterson, A. H., Bowers, J. E., Bruggmann, R., Dubchak, I., Grimwood, J., Gundlach, H. and Rokhsar, D. S. 2009. The Sorghum bicolor genome and the diversification of grasses. Nature, 457(7229): 551–556.

- Santos, M., Romano, E., Yotoko, K., Tinoco, M., Dias, B. and Argao, F. 2005. Characterisation of the Cacao Somatic Embryogenesis Receptor-Like Kinase (*SERK*) Gene Expressed during Somatic Embryogenesis. *Plant Sci.*, 168:723-729
- Schmidt Ed, D.L., Guzzo, F., Toonen, M.A.J. and de Vries, SC. 1997. A leucine–rich repeat containing receptor-like kinase marks somatic plant cells competent to form embryos. *Development*, 124:2049–2062.
- Singla, B., Khurana, J.P. and Khurana, P. 2008. Characterization of three somatic embryogenesis genes from wheat, *Triticum aestivum*. *Plant Cell Rep.*, 27:833-843
- Somaleva, M.N., Schmidt, E.D.L. and de Vries, S. 2000. Embryogenic cells in *Dactylis glomerata* L. (Poaceae) explants identified by cell tracking and by *SERK* expression. *Plant Cell Rep.*, 19:718–726.
- Stone, S.L., Kwong, L.W., Yee, K.M., Pelletier, J., Lepiniec, L., Fischer, R.L., Goldberg, R.B. and Harada, J.J. 2001. LEAFY COTYLEDON2 encodes a B3 domain transcription factor that induces embryo development. Proc Natl Acad Sci USA, 98: 11806–11811
- Thomas, C., Meyer, D., Himber, C. and Steinmetz, A. 2004. Spatial expression of a sunflower *SERK* gene during induction of somatic embryogenesis and shoot organogenesis. *Plant Physiol Biochem.*, 42:35-42.
- Varner, J.E. and Lin, L.S.1989. Plant cell wall architecture. *Cell*, 56:231-239.
- Zuo, J.R., Niu, Q.W., Frugis, G. and Chua, N.H. 2002. The *WUSCHEL* gene promotes vegetative-to-embryonic transition in Arabidopsis. *Plant J.*, 30, 349–359.
