



RESEARCH ARTICLE

ANTIFUNGAL ACTIVITY OF APPLE CIDER VINEGAR AGAINST CLINICAL ISOLATES OF CANDIDA SPECIES

<sup>1</sup>Janhvi Manohar and <sup>2,\*</sup>Dr. Gopinath, P.

<sup>1</sup>BDS 2nd year, Saveetha Dental College, Chennai, India

<sup>2</sup>Senior Lecturer, Department of Microbiology, Saveetha Dental College, Chennai, India

ARTICLE INFO

Article History:

Received 16<sup>th</sup> January, 2017  
Received in revised form  
15<sup>th</sup> February, 2017  
Accepted 25<sup>th</sup> March, 2017  
Published online 30<sup>th</sup> April, 2017

Key words:

Candida spp,  
MIC,  
Apple Cider Vinegar.

ABSTRACT

Cutaneous fungal infections are common diseases in humans, and can also be caused by dermatophytic fungi and some yeasts. Superficial candidiasis is a common infection of the oral cavity, vagina, skin and esophagus, although most infections occur in patients who are immunocompromised. *Candida albicans* responsible for many of these infections, but occasionally other members of the genus are associated, and generally infect the skin, nails, or mucous membranes. This study involves the determination of antifungal activity of *Candida* spp against apple cider vinegar. The MIC of apple cider vinegar was appeared to be 0.25% for *Candida* spp. It is also a safer treatment modality with almost no side-effects. Apple cider vinegar seems to be the most economical product to have antifungal properties. Such activity may suggest promising effective alternative or synergistic remedy to the popular antifungal drugs.

Copyright©2017, Janhvi Manohar and Dr. Gopinath. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Citation: Janhvi Manohar and Dr. Gopinath. 2017. "Antifungal activity of apple cider vinegar against clinical isolates of *Candida* species", *International Journal of Current Research*, 9, (04), 49317-49320.

INTRODUCTION

Cutaneous fungal infections are common diseases in humans, and can also be caused by dermatophytic fungi and some yeasts. Superficial candidiasis is a common infection of the oral cavity, vagina, skin and esophagus, although most infections occur in patients who are immunocompromised. *Candida albicans* responsible for many of these infections, but occasionally other members of the genus are associated, and generally infect the skin, nails, or mucous membranes (Hall, 1999; Lacaz, 2002; Eggimann, 2003; Hay, 2005). The cause for vulvovaginal candidiasis is usually *C. albicans* affecting almost 95% of women followed by *C. glabrata* affecting 10-20% along with *C. tropicalis* and *C. krusei*. (Sobel, 2007; Develoux, 2007). The candida species are known to be gram-positive yeast-like fungi that take up an oval or round shape. They can be isolated more commonly from primary endodontic infections, plaque, caries, subgingival microflora and active periodontal cavity. *C. albicans* are also a part of the normal oral microflora (Radeva, 2007) *C. albicans* is found to be the most common fungus involved in endodontic re-infections. The root canal is disinfected through mechanical preparation, chemical irrigation and placement of intracanal medicaments (Abbasi, 2015).

The conventional treatment of fungal disease is limited, and part of the reason is due to the limited spectrum of the currently antifungal drugs, and the expensive treatment, particularly due to the need of prolonged therapy. Thus, new drugs and alternative therapies are necessary, including natural products. (Silva, 2008). In recent studies, essential oils and extracts derived from natural plant species have been known to produce anti-microbial property against virulent genus of microorganisms. Investigations concerning the evaluation of the biological activities of essential oils of some medicinal plants have revealed that some of them exhibited antibacterial, antifungal and insecticidal properties (Burt, 2004). Because of the antimicrobial properties showed by essential oils, the aromatherapy has been used for treatment of serious skin diseases, in special, superficial mycoses (Carlini, 1986) Similarly, in this study the anti-fungal activity of apple cider vinegar was assessed. Apple cider vinegar (ACV) is a solution of acidic acid produced by fermentation of apples. Scientists have measured ninety different substances in apple cider vinegar such as thirteen types of carboxylic acids, four aldehydes, twenty ketones, eighteen types of alcohols, eight ethyl acetates etc. It also contains important minerals, trace elements and vitamins (as listed underneath) as well acetic acid, propionic acid, lactic acid and malic acid, enzymes, amino acids as well as roughage in the form of potash and apple pectin. (Jabir, 2011) ACV has various benefits like eliminating bad cholesterol, weight reduction, anti-aging and most

\*Corresponding author: Dr. Gopinath, P.,  
Senior Lecturer, Department of Microbiology, Saveetha Dental College, Chennai, India.

importantly has anti-fungal, anti-bacterial, anti-viral and anti-cancer effects. However, the FDA has not recognised ACV as a potent anti-microbial as there isn't scientific evidence for it. This study aims to create awareness among people about the various benefits of ACV, a household ingredient.

**MATERIALS AND METHODS**

**CANDIDA ISOLATES**

A total of 20 non-repetitive clinical isolates of *Candida species* were collected from different samples of immunocompromised individuals attending Saveetha Medical college, Thandalam. They were characterized by carbohydrate fermentation and assimilation tests and confirmed (Salkin, 2000). Isolates were preserved in semisolid Sabouraud chloramphenicol semi solid stock and stored at 4°C until further use.

**CHARACTERIZATION OF CANDIDA SPECIES**

*Candida* species were further characterized by using Hichrom agar (Himedia, Mumbai).

**Preparation of Hichrom agar**

CHROMagar *Candida* (HiMedia, Mumbai) was prepared following manufacturer's instructions. About 21.02 gram of HiChrome *Candida* differentiation agar base (modified) was suspended in 500 ml of distilled water. It was heated to boiling gently to dissolve the medium completely. Then it was allowed to cool to 50°C and rehydrated (one vial) contents of Hichrome *Candida* selective supplement was added under aseptic precautions. It was mixed well and poured into petri dishes. Isolates were identified on Hichrome agar based upon the characteristic color of the colony by sub culturing from Sabouraud's chloramphenicol agar plates and the *Candida* Hichrome plates were incubated at 37°C for 24- 48 hours (Abbasi, 2015). Based on colour produced by the isolates speciation have been made.

<i>Candida</i> species	Colour
<i>C.albicans</i>	Green
<i>C.tropicalis</i>	Blue
<i>C.krusei</i>	Pink dry colonies
<i>C.kefyr</i>	Pale
<i>C.parapsilosis</i>	Pale

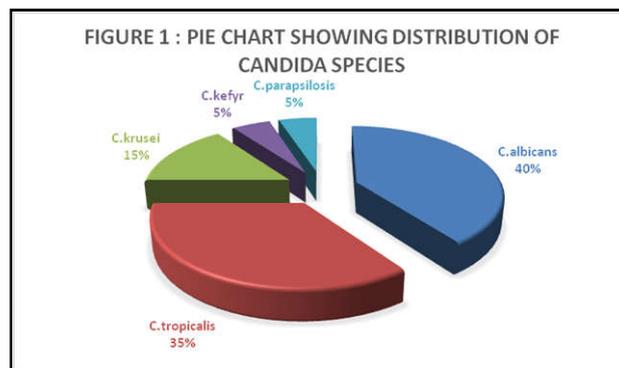
**Detection of antifungal activity of apple cider vinegar against clinical isolates of *Candida* species**

Antifungal activity of apple cider vinegar was tested against *Candida* isolates by minimum inhibitory concentration method. Agar dilution method was performed to attain the different concentrations of apple cider vinegar such as 0.03%, 0.06%, 0.125%, 0.25%, 0.5%, 1% and 2% SDA. Media containing various concentrations of vinegar were poured over the sterile petri dishes and allowed to dry. Media without vinegar was served as control plate. Spot inoculation of 0.5 McFarland standard turbidity adjusted isolates were made on the plates and incubated at 37°C for overnight. The lowest concentration of the essential oils that completely inhibited the growth of isolates was considered as MIC.

**RESULTS**

**Characterization of *Candida* species by Hichrom *Candida* agar**

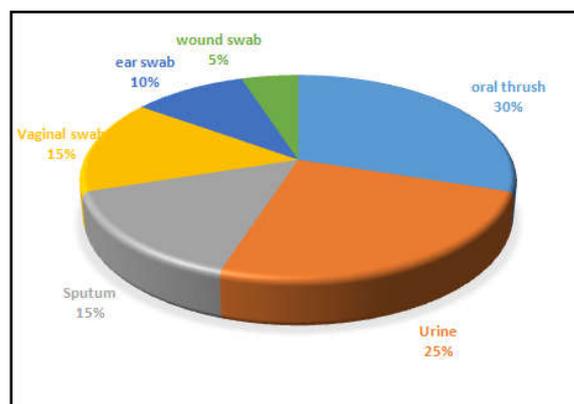
A total of 20 clinical isolates of *Candida* spp were seeded on to Hichrom *Candida* agar and results were tabulated based on pigment production.



Representative picture showing *Candida* species on Hichrom *Candida* agar

**Sample wise distribution of *Candida* species**

Of the 20 clinical isolates of *Candida*spp, 6/20 (30%) were from oral thrush, 5/20 (25%) from urine, 3/20 (15%) from sputum, 3/20 (15%) from vaginal swab, 2/20 (10%) from ear swab and one (5%) from wound swab.



Pie chart showing the sample wise distribution of *Candida* species

**Result of antifungal activity of apple cider vinegar against clinical isolates of *Candida* species**

We have observed that, clinical isolates of *Candida* spp were inhibited from 0.25-0.5% of apple cider vinegar. The MIC of apple cider vinegar was appeared to be 0.25% for *Candida* spp.

Dilutions of apple cider vinegar	0.03%	0.06%	0.125%	0.25%	0.5%	1%	2%
No. of organisms	0	0	0	18 (90)	2 (10)	0	0



## DISCUSSION

Vinegar, alone, has been used for cleaning and treating nail fungus, head lice, warts and ear infections. Consumers typically prefer natural preservative methods for inhibiting the growth of food borne pathogenic microorganisms in food. (Rauha *et al.*, 2000) Several problems might be encountered with antifungal drugs, first; resistance; as fungi may become resistant to antifungal drugs due to target gene mutations, enzyme modification or to development of pump system that expels the drug out of the fungal cell (Finkel *et al.*, 2005) second; toxicity; as antifungal drugs can cause systemic toxicities as hepatotoxicity and nephrotoxicity. These problems necessitate searching for safer effective remedies with known antifungal activity (Jabir, 2011).

Apple cider vinegar is a commonly prescribed antifungal agent in folk medicine for treatment of fungal skin, ear and vaginal infections. (Jabir, 2011) The antifungal activity of apple cider vinegar might be attributed to its malic acid, acetic acid contents or to other non-identified ingredients. The mechanism of inhibition of fungal growth by organic acids is generally not considered a pH phenomenon. It is known that, growth and morphology of fungi are influenced by the pH of media (Higgins, 1999). Organic acids resulting in a decreasing pH value, this may influence the growth by acidifying the cell, which will consume a great amount of energy to maintain the intracellular pH homeostasis (Kang *et al.*, 2003). Other explanations have also been proposed including the membrane disruption, the interruption of metabolic reactions, and the accumulation of toxic anions (Hassan *et al.*, 2015).

The inhibition of microbial growth increases by lowering pH of the media, and most microorganisms are susceptible to antimicrobial effects in the presence of organic acids. This phenomenon is due to the hydrophobic feature of most organic acids, which allows free diffusion of the protonized form through cell membrane. This diffusion process takes place spontaneously due to pH and osmolarity gradients that exist between the inner and outer sides of the cell. The intracellular pH is higher than the extracellular, and the acid undergoes dissociation as soon as it enters the cytoplasm and then decreases the intracellular pH by releasing the proton. To counter the decrease of cytoplasmic pH, resulting from the ionization of the entered acid, the cell allocates the main part of its energy content to eliminate these newly formed protons which results in slower growth kinetics (Pelaez *et al.*, 2012) According to Hassan. R *et al.* (Pelaez *et al.*, 2012), acetic acid shows strongest inhibition of fungal growth among other organic acids. Similarly, the study done by Prakasam *et al.* stated that natural products like aloe vera showed better inhibition when compared to calcium hydroxide. In this study,

the MIC of apple cider vinegar against various isolates of *Candida* species appeared to be 0.25% for *Candida* spp. Antifungal activity is seen between 0.25% to 0.5% dilute concentrations of ACV. Thus, less than 1% of ACV is sufficient to produce anti-fungal effect against *Candida*.

## Conclusion

*Candida* is one of the commonest opportunistic pathogens that cause disease in compromised hosts. (Nivedhaa Chezhan, 2015) Natural oils and products seem to be more efficient in their anti-fungal property when compared to anti-fungal drugs. It is also a safer treatment modality with almost no side-effects. Apple cider vinegar seems to be the most economical product to have anti-fungal properties. Such activity may suggest promising effective alternative or synergistic remedy to the popular antifungal drugs.

## REFERENCES

- Abbasi, M., Norouzigard, A., Sharifi, M. 2015. Journal of Islamic Dental Association of Iran (JIDAI) Winter, 26, 4.
- Burt, S. 2004. Essential oils: their antibacterial properties and potential applications in foods—a review. *Int J Food Microbiol*, 94:223-53.
- Carlini, E.A., Contar, J.D.P., Silva-Filho, A.R. *et al.* Pharmacology of lemongrass (*Cymbopogon citratus* Stapf). I. Effects of tea prepared from the leaves on laboratory animals. *J Ethnopharmacol* 1986;17:37-64.
- Develoux, M., Bretagne, S. 2005. Candidosis et levures diverses. *EMC Maladies Infectieuses*, 2:119-39.
- E Radeva; B Indjov; R Vacheva, *Journal of IMAB*, 2007; 13, 2.
- Eggimann, P., Garbino, J., Pittet, D. 2003. Epidemiology of *Candida* species infections in critically ill non-immunosuppressed patients. *Lancet Infect Dis.*, 3:685-702.
- Finkel, Richard; Clark, Michelle A., Cubeddu, Luigi X. 2009. Lippincott's Illustrated Reviews: Pharmacology, 4th Edition.
- Hall J.C. 1999. Sauer's manual of skin diseases. 8. ed. Philadelphia: Lippincott Williams & Wilkins.
- Hassan, R., El-Kadi, S.S. and M. 2015. Effect of some organic acids on some fungal growth and their toxins production. *International Journal of Advances in Biology (IJAB)* Vol 2. No .1, February, 1-11
- Hay, R. 2005. Superficial fungal infections. *Skin and Soft Tissue Infections*, 33:89-90.
- Higgins, C. and Brinkhaus, F. 1999. Efficacy of several organic acids against molds, *J. Appl. Poultry Res.*, Vol. 8, pp 480-487.
- Jabir, H. *et al.* 2011. In vitro assessment of antifungal potential of apple cider vinegar and acetic acid versus fluconazole in clinical isolates of otomycosis. *Thi-Qar Medical Journal (TQMJ)*: Vol (5) No(1): (126-133).
- Kang, H. C., Park, Y. H. and Go, S. J. 2003. Growth inhibition of a phytopathogenic fungus, *Colletotrichum* species by acetic acid, *Microbiol. Res.* Vol. 158, pp 321–326.
- Lacaz C.S., Porto E., Martins J.E.C. *et al.* 2002. Tratado de micologia médica, São Paulo, Sarvier.
- Nivedhaa Chezhan *et al* /J. Pharm. Sci. & Res. Vol. 7(9), 2015, 767-768
- Pelaez, A. M. L., C. A. S. Catano, E. A. Q. Yepes, R. R. G. Villarroel, G. L. D. Antoni and L. Giannuzzi, 2012. Inhibitory activity of lactic and acetic acid on *Aspergillus*

- flavus growth for food preservation. *Food Control*, Vol. 24, pp 177-183.
- Rauha, J.P., Remes, S., Heinonen, M., Hopia, A., Kahkonen M., Kujala, T., Pihlaja, K., Vuorela, H., Vuorela, P. 2000. Antimicrobial effect of Finnish plant extracts containing flavonoids and other phenolic compounds. *Intl J Food Microbiol* 56:3-12.
- Ravikumar, C., Chandana, C.S., Prakasam, G. Comparative study on the intracranial medicaments based on its antifungal efficacy. *World Journal of Pharmacy and Pharmaceutical Sciences*, Vol 5, 11, pp 1523-1527
- Salkin, I.F., Pruitt, W.R., Padhye, A.A., Sullivan, D., Coleman, Pincus. 2000. Carbohydrate assimilation profiles used to identify first clinical isolates of *Candida dubliniensis* in United States. *J Clin Microbiol*, 36:1467.
- Silva, C. D., Guterres, S. S., Weisheimer, V., & Schapoval, E. E. 2008. Antifungal activity of the lemongrass oil and citral against *Candida* spp. *Brazilian Journal of Infectious Diseases*, 12(1).
- Sobel, J.D. 2007. Vulvovaginal candidosis. *Lancet*, 369:1961-71

\*\*\*\*\*