

Available online at http://www.journalcra.com

International Journal of Current Research Vol. 11, Issue, 06, pp.4321-4327, June, 2019 INTERNATIONAL JOURNAL OF CURRENT RESEARCH

DOI: https://doi.org/10.24941/ijcr.35654.06.2019

RESEARCH ARTICLE

COMPARATIVE STUDIES ON THE ANTIMICROBIAL ACTIVITY OF WATERY LEAFY EXTRACTS FROM LAWSONIA INERMIS L.(HENNA), OROXYLUM INDIUM (L.)VENT (MIDNIGHT HORROR) AND MELASTOMA MELABATHRICUM L. (MALABAR)

¹Dr. Kyaw Zan Aung and ^{2, *}Dr. Mi Mi Yee

¹Professor and head of Chemistry Department, Panglong University ²Associate Professor of Chemistry Department, Panglong University

ARTICLEINFO

ABSTRACT

Article History: Received 10th March, 2019 Received in revised form 19th April, 2019 Accepted 20th May, 2019 Published online 30th June, 2019

Key Words: Lawsonia inermis L., Antimicrobial activity and Phytochemical Screening.

In the present investigation the antimicrobial activities of some traditional medicinal plants which were selected based on medicinal reports practiced by native people of Myeik Township, Tanintharyi region in Myanmar. Antimicrobial activities of aqueous extract of Lawsonia inermis L.(Henna), Oroxylum indium (L.)Vent (midnight horror) and Melastoma melabathricum L.(Malabar) tested against the following microorganism like Staphylococcus aureus, Bacillus pumilus, Bacillus subtilis, Pseudomonas aeruginosa , Escherichia coli and Candida albicans by Agar-well diffusion method. The Lawsonia inermis L. showed maximum zone of inhibition against Staphylococcus aureus, (15 mm) and minimum (13 mm) of Candida albicans. The Oroxylum indium (L.)Vent showed maximum zone of inhibition against Staphylococcus aureus, (13 mm) and minimum (11 mm) of Pseudomonas aeruginosa. Melastoma melabathricum L. showed maximum zone of inhibition against Bacillus subtilis (13 mm) and the minimum (12 mm) of Candida albicans. The results revealed the presence of important medicinal phytochemicals constituents, such as alkaloids, flavonoids, phenols, saponins, α amino acids, reducing sugar, phlobatannins and tannins. were present in the Lawsonia inermis L. and Melastoma melabathricum L.For Oroxylum indium (L.)Vent, presence of alkaloids, glycosides, phlobatannins, phenolic compounds, α -amino acids, saponins and tannins which interpret its medicinal values. Among the three samples, coumarins was found to be presence in Lawsonia inermis L

*Corresponding author: Dr. Mi Mi Yee

Copyright © 2019, Kyaw Zan Aung and Mi Mi Yee. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Citation: Dr. Kyaw Zan Aung and Dr. Mi Mi Yee, 2019. "Comparative Studies on the Antimicrobial Activity of Watery Leafy Extracts from Lawsonia inermis L. (Henna), *Oroxylum indium* (L.)Vent (midnight horror) and *Melastoma melabathricum L.* (Malabar)", *International Journal of Current Research*, 11, (03), 4321-4327.

INTRODUCTION

Medicinal plants have been a major source of treatment for human diseases since time immemorial. One fourth of the world population i.e. 1.42 billion people are dependent on traditional medicines, particularly plant drug for curing ailments Herbal medicines are promising choice over modern synthetic drugs. They show minimum side effects and are considered to be safe (9). In this research work, the aqueous extract of Lawsonia inermis L.(Henna) leaves, Oroxylum indium (L.)Vent (midnight horror)leaves and Melastoma melabathricum L. (Malabar) leaves from Tanintharyi region in Myanmar which were used to traditional medicines of wound healing and hypertension treatment, were checked its authenticity and purity as per the standard parameter. Lawsonia inermis, also known as henna, the henna tree, the mignonette tree, and the Egyptian privet, (Bailey, 1976) is a flowering plant and the sole species of the Lawsonia genus. It is the source of the dye Dan used to dye skin, hair and fingernails, as well as fabrics including silk, wool and

Leather (Okwu, 2006). The henna plant is native to northern Africa, western and southern Asia, and northern Australasia, in semi-arid zones and tropical areas. It produces the most dye when grown in temperatures between 35°C and 45 °C (95 and 113 °F) (Leal, 2000). Main chemical constituents of the Henna are Lawsone (2-hydroxynaphthoquinone), mucilage, mannite, gallic acid and tannic acid (Rubiay, 2008). Henna is known to be used as cosmetic agent for dyeing hair, nails and skin(20).In traditional medicine, Henna plant is used to treat many diseases like oedema, bronchitis, menstrual disorder, rheumatism, hemorrhoids and even injaundice, leprosy, pain, spleenen largement, dysentery and skin problems (Cuong, 2009). Henna can also be used as anastringent and antihemorragic agent and is also known for its hypotensive, cardio inhibitory and sedative effects (Warrier, 1995) .In addition, henna is reported to show some other properties including hypoglycemic immstimulant, hepatoprotective, antiinflammatory, tuberculostatic ,anticancer and antioxidant properties (Syamsudin, 2008).

Oroxylum indicum. (L.) (midnight horror) decoction is given in treating rheumatic pain, enlarged spleen (Khare, 2004), ulcer, cough, and bronchitis. Mature Fruits are acrid, sweet, anthelmintic, and stomachic. They are useful in pharyngodynia, cardiac disorders, gastropathy, bronchitis, haemarrhoids, cough, piles, jaundice, dyspepsia, smallpox, leucoderma and cholera (Kala, 2011). Seeds are used as purgative. Dried seed powder is used by women to induce conception. Seeds yield non-drying oil used in perfume industry. The seeds are ground with fire soot and the paste is applied to the neck for quick relief of tonsil pain. The seeds are used in traditional Indian Ayurvedic medicine, included in famous tonic formulations such as Chyawanprash (Olaleye, 2007). Bark decoction is taken for curing gastric ulcer and a paste made of the bark powder is applied for mouth cancer, scabies and other skin diseases. The medicated oil of O. indicum in sesame oil base instilled into ears mitigates the pain in otitis (Warrier, 1995) and fiber. Roots are sweet, astringent, bitter, acrid, refrigerant (Yoganarasimhan, 1996) antiinflammatory, anodyne, aphrodisiac, expectorant, appetizer, carminative, digestive, anthelmintic, constipating, diaphoretic, diuretic, antiarthritic, antidiabetic and febrifuges. Tonic is useful in dropsy, cough, sprains neuralgia, hiccough, asthma, bronchitis, anorexia, dyspepsia, flatulence, colic, diarrhea, dysentery, strangury, gout, vomiting, leucoderma, wounds, rheumatoid arthritis and fever. Root bark is used in stomatitis. nasopharyngeal cancer and tuberculosis (Bhattacharje, 2005). Leaves are used as stomachic, carminative and flatulent. Several workers have reported different biochemical activities of O. indicum in various in vivo and in vitro test models. Different part of this plant have been found to exhibit anti-inflammatory, antimicrobial, antioxidant, anticancer, anti-mutagenic, photocytotoxic, antiarthritic, immunostimulant, hepatoprotective, anti-proliferative and hepatoprotective activities.

Melastomataceae (Malabar) plants originate in the tropic and subtropic regions, with a total of more than 4000 species in the world. In the Southeast Asian region alone, the genus Melastoma comprises 22 species, 2 subspecies, and 3 varieties (Rajenderan, 2010). In general, M. malabathricum is a small shrub commonly found in previously cleared land, waste places, and roadside throughout the Southeast Asian countries, including Tanintharyi region in Myanmar. It is native to tropical and temperate Asia and the Pacific Islands (Ling, 2009). The plant is one of the most common weeds that grow wildly and abundantly throughout the tropics, especially in the moist areas, and can be found in the Indian Ocean Islands, throughout South and South-East Asia, China, Taiwan, Australia, and the South Pacific Ocean (Wong, 2008). The leaves are chewed up, pounded, and applied as paste on cuts or wounds or finely chopped up and squeezed to apply the juice onto the wound to stop bleeding (Latiff, 2000). According to Sharma et al., the leaves can also be used to prevent scarring from smallpox, to treat dysentery, diarrhea, and piles, and as a tonic (Zakaria et al., 2006). The young leaves are eaten to treat diarrhea while the young premature leaves are consumed raw to cure dysentery (Koay, 2008). The shoots can be ingested to treat puerperal infections, high blood pressure, and diabetes while the shoots juice can also be used as a mouthwash to relieve a toothache or to treat leukorrhea. Other than those mentioned above, the leaves are also medicinally useful to treat ulcers, gastric ulcers, scar, pimple, and black spot at skin. The roots can also be used as mouthwash to relieve a toothache and to treat epilepsy (Burkill, 1996) given to postpartum women to

aid healing and womb strengthening (Fazlin, 2002) or to alleviate rheumatism, arthritis, and tenderness in the legs (Koay, 2008) Other than that, the powdered leaves and roots can be applied to wounds and pox scars to aid the healing process (Fazlin, 2002) or used to relieve the discomfort of hemorrhoids with the former also used as astringent for dysentery (Sajem, 2006), (Strasser, 2010).

MATERIALS AND METHODS

Sample Collection: Fresh Leaves of *Lawsonia inermis* L.(Henna), *Oroxylum indium* (L.)Vent(midnight horror) and *Melastoma melabathricum* L.(Malabar)were collected from Myeik Township, Thanintharyi Division, Myanmar. The collected sample was identified in Department of Botany, University of Myeik.

Chemicals: All chemicals used in this work were from British Drug House Chemical Ltd., Poole, England. All standard solutions and other diluted solutions throughout the experimental runs were prepared by using distilled water. In all the investigations the recommended methods and standard procedures involving both conventional and modern techniques were employed (Vogel, 1978). All other chemicals and reagents used were of analytical grae.

Preparation of leaf extracts: The sample was cleaned from dust, washed with water, chopped, and dried at room temperature for one week. The dried material was made powder by using grinding machine, and stored in airtight glass bottle until used. The dry powdered sample material (15)g was extracted with 100ml of water ,ethanol, and methanol separately .The contents were kept as such in room temperature for 48h with constant stirring at regular intervals After the incubation period, the contents were filtered through Whatman No.1 filter paper. Then filtrate were vacuum dried using rotary evaporator and concentrates were stored at 4°C.The residues were redissolved with the appropriate solvents from which they were prepared and used for further studies.

Preliminary Phytochemical analysis: Qualitative phytochemical analyses were performed in filtrates of *Lawsonia inermis* L. (Henna), *Oroxylum indium* (L.)Vent (midnight horror) and *Melastoma melabathricum* L. (Malabar) preliminary phytochemical test were carried out according to determine the prescence of phytochemicals the alkaloid, α --amino acids, carbohydrates ,flavonoids, phenolic compounds, reducing sugar, tannin ,starch ,glycosides, saponins , coumarins and phlobatannins as described by standard procedure.

Elemental analysis: Elemental analysis of *Lawsonia inermis* L.(Henna), *Oroxylum indium* (L.)Vent (midnight horror) and *Melastoma melabathricum* L. (Malabar) samples were determined by EDXRF method.

Test organism: Screening of Antimicrobial activity of *Lawsonia inermis L.* (Henna), *Oroxylum indium (L.)Vent* (midnight horror) and Melastoma melabathricum L. (Malabar) samples were determined by agar well diffusion method in DCPT, Yangon. Six species of microorganisms such as *Bacillus subtilis, Staphylococcus aureus, Pseudomonas aeruginosa, Bacillus pumilus, Candida albican* and *Escherichia coli* were treated with the sample.

Preparation of inoculums: The microorganisms were inoculated into nutrient broth and rose Bengal broth for bioassay and incubated For 24 and 48 h at 37°C. The turbidity of the medium indicates the growth of organisms.

Antimicrobial studies: The agar well diffusion method was employed for the determination of antimicrobial activity of extracts. Lawn culture of *E.coli, Candida albican, Bacillus puimilus, psrudomonus aeruginosa, Staphylococcus aureus and Bacillus subtils* were spread on nutrient agar and A. niger & A flavus spread on rose bengal agar using sterile cotton swabs. The wells (6mm in diameter) were cut from the agar plates using a cork horer.30µlof the extracts (7mg/ml)were poured into the well using a sterile micro pipette .The plates were incubated at $37\pm2^{\circ}$ C for 24 hours for bacterial activity and 48 hours for fungal activity .The zone of inhibition was calculated by measuring the diameter of the inhibition zone around the well (in mm) including the well diameter.

RESULTS AND DISCUSSION

In this research work, the aqueous extract of *Lawsonia inermis* L.(Henna) , *Oroxylum indium* (L.)Vent (midnight horror)and *Melastoma melabathricum* L.(Malabar) sample leaves from Tanintharyi region in Myanmar which were used to traditional medicines of wound healing and hypertension treatment, were checked its authenticity and purity as per the standard parameter.



Figure 1. The plant and leaves of *Oroxylum indicum*. (L.)Vent(midnight horror)



Figure 2. The plant and leaves of *Lawsonia inermis* L. (Henna)

Preliminary phytochemical profiling reflects essential information regarding the diversity of different classes of secondary metabolites such as alkaloids, flavonoids, steroids, saponins, tannins, reducing sugars etc. in the plant extracts.



Figure 3. The plant and leaves of *Melastoma melabathricum* L. (Malabar)

In the present study, qualitative tests for all three crude extracts showed significant indication about the presence of various secondary metabolites. Preliminary phytochemical screening of Lawsonia inermis L, Oroxylum indium (L.)Vent and Melastoma melabathricum L.leaves extracts revealed the presence of various bioactive compounds and the results were presented in Table 1. The phytochemical results of Lawsonia inermis L.leaves extracts indicated positive for all except starch while Oroxylum indium (L.)Vent extracts tested positive for all except flavonoids, and coumarins. The phytochemical results of Melastoma melabathricum L. leaves extracts indicated positive for all except starch and coumarins. The people of Myeik in Tanintharyi Region also used the fresh and dry leaves of Lawsonia inermis L. and Melastoma melabathricum L. to treat anti-inflammatory and cuts and wounds.

This is due to the presence of saponin, flavonoids tannins and phlobatannins in Lawsonia inermis L leaves and Melastoma melabathricum L.leaves. The presence of saponins, and tannins were also reason why the leaves are used traditionally to use wound healing (Malinow, 1977). The juice from the Lawsonia inermis L. leaves and Melastoma melabathricum L. leaves have been found to be anti-inflammatory .This can be explained due to the presence of saponins and flavonoids as both constituent show anti-inflammatory properties (Kenner, 1996). The presence of phlobatannins in Lawsonia inermis L leaves and Melastoma melabathricum L. leaves which have been reported for its wound healing properties, these are antiinflammatory and analgesic (Ayinde, 2007) and antioxidant (Okwu, 2004). Moreover, the presence of alkaloids and saponins in the leaves of Oroxylum indium (L.)Vent as shown in Table explains why the leaves of Oroxylum indium (L.)Vent used for hypertension treatment, mainly in Myeik Township, Tanintharyi Region (Akinpelu, 2006). Among the three samples, coumarins was found to be in Lawsonia inermis L. Many pharmacological activities have been ascribed to coumarins such as anticlotting, hypotensive, antimicrobial, anti-inflammatory, and antitumor activities .The results revealed the presence of important medicinal phytochemicals constituents, such as terpenoids, reducing sugar, flavonoids, alkaloids and phlobatannins were present in the Lawsonia inermis L. and Melastoma melabathricum L. For Oroxylum indium (L.) Vent, presence of alkaloids, glycosides, phlobatannins, phenolic compounds, α -amino acids, saponins and tannins which interpret its medicinal values. EDXRF spectrometer was used to verify the relative quantitative percentage of elements in the leaves of Lawsonia inermis L., Oroxylum indium (L.)Vent and Melastoma melabathricum L.

4324 Kyaw Zan Aung and Mi Mi Yee, Comparative Studies on the Antimicrobial Activity of Watery Leafy Extracts from Lawsonia inermis L. (Henna), Oroxylum indium (L.)Vent (midnight horror) and Melastoma melabathricum L. (Malabar)



Figure 4. EDXRF spectrums of Lawsonia inermis L. (henna) leaf



Figure 5. EDXRF spectrums of Oroxylum indium (L.)Vent (midnight horror) leaf



Figure 6. EDXRF spectrums of Melastoma melabathricum L. (Malabar) leaf

No	Tests	Henna Leaves	midnight horror Leaves	Malabar Leaves
1.	Alkaloids	+	+	+
2.	Glycosides	+	+	+
3.	Flavonoids	+		+
4.	Carbohydrates	+	_ +	+
5.	Phenolic Compounds	+	+	+
6.	-amino acids	+	+	+
7.	Saponins	+	+	+
8.	Starch	-	+	-
9.	Tannins	+	+	+
10.	Reducing sugar	+	+	+
11	coumarins	+		
12	phlobatannins	+	+	+

Table 1. Summary of the Phytochemical Investigation Results of Three Selected Leaves

 Table 2 Element Content in Lawsonia inermis L, Oroxylum indium (L.) Vent and Melastoma melabathricu L.

Element	Quantitative Results (%)					
	Lawsonia inermis L.	Oroxylum indium (L.)Vent	Melastoma melabathricu L.			
~		1 (0)				
Ca	1.071	1.696	1.605			
K	1.034	1.323	0.642			
S	0.335	0.358	0.461			
Р	0.051	0.084	ND			
Fe	0.016	0.015	0.010			
Mn	0005	0.003	0.036			
Br	0.002	ND	0.001			
Zn	0.002	0.009	0.002			
Sr	0.002	ND	0.002			
Cu	0.002	0.005	0.001			
Rb	0.000	0.001	0.001			
Os	ND	0.001	ND			
Ti	ND	ND	0.003			
COH	97.481	96.505	97.038			

N D =Non detective

 Table 3. Inhibition Zone (mm) of Antimicrobial Activity for Aqueous Extract from Leaves of Lawsonia inermis L.

 Oroxylum indium (L.)Vent and Melastoma melabathricum L.

Compound	Zone of inhibition (mm)							
	Gram- positive bacteria			Gram- negative bacteria		Fungi		
	B. Sub	S.aureus	B.pumils	Pseudomonas	E.coli	Candida		
Lausonia incomis I	14mm	15mm	14mm	13mm	13mm	13mm		
Lawsonia inermis L.	(+)	(++)	(+)	(+)	(+)	(+)		
Oroxylum indium(L.)Vent	13mm	13mm	13mm	11mm(+)	12mm	12mm		
	(+)	(+)	(+)		(+)	(+)		
Melastoma melabathricum	13mm	12mm	12mm	12mm	13mm	12mm		
L.	(+)	(+)	(+)	(+)	(+)	(+)		
A 11 10	*0 *							

Agar well – 10mm <u>*Organisms*</u>

10mm ~ 14mm (+) Bacillus sublilis (N.C.T.C-8236), Bacillus pumilus (N.C.I.B- 8982)

15mm ~ 19mm (++) Staphylococcus aureus (N.C.P.C-6371), Candida albican

20mm above (+++) Pseudomonas aeruginosa (6749), E-coli (N.C.I.B -8134)

The present amount of elements present in Lawsonia inermis L, Oroxylum indium (L.)Vent and Melastoma melabathricum L. was shown in Table 2 and figure 4,5 and 6.The antimicrobial activity of the aqueous extracts of three different leaves were also determined by Agar well diffusion method The antibacterial and antifungal activities of aqueous extract from leaves, such as Lawsonia inermis L. Oroxylum indium (L.)Vent and Melastoma melabathricum L.tested against the following microorganism like Staphylococcus aureus, Bacillus pumilus and Bacillus subtilis as gram-positive bacteria, Pseudomonas aeruginosa and Escherichia coli as gramnegative bacteria and Candida albicans as fungal strain by Agar-diffusion method. The results indicated that Lawsonia inermis L.has shown the maximum antibacterial and antifungal activities against all tested microorganism as shown in Table 3 and figure 7.

The Lawsonia inermis L. showed maximum zone of inhibition against Staphylococcus aureus, (15 mm) followed by Bacillus subtilis (14 mm), Bacillus pumilus (14 mm), Pseudomonas aeruginosa (13 mm), Escherichia coli (13mm) and Candida albicans (13 mm). The Oroxylum indium (L.)Vent showed maximum zone of inhibition against Staphylococcus aureus, (13 mm) followed by Bacillus subtilis (13 mm), Bacillus pumilus (13 mm), Pseudomonas aeruginosa (11 mm), Escherichia coli (12mm) and Candida albicans (12 mm). Melastoma melabathricum L. showed maximum zone of inhibition against Staphylococcus aureus, (12mm) followed by Bacillus subtilis (13 mm), Bacillus pumilus (12 mm), Pseudomonas aeruginosa (12 mm), Escherichia coli (13mm) and Candida albicans (12 mm). The overall observation of antimicrobial activity of aqueous extract from three leaves indicated that the Lawsonia inermis L. have more impact than 4326

the *Oroxylum indium* (L.)Vent and *Melastoma melabathricum* L. This is due to the presence of coumarins in Lawsonia inermis. Coumarins have many pharmacological activities such as anticlotting, hypotensive, antimicrobial, anti-inflammatory, and antitumor activities .Among the *Oroxylum indium* (L.)Vent and *Melastoma melabathricum* L. *Oroxylum indium* (L.)Vent has shown the moderate against the gram- positive bacteria than the *Melastoma melabathricum* L.

Conclusion

From the result it can be concluded that the aqueous extract from leaves of Lawsonia inermis L., Oroxylum indium (L.)Vent and Melastoma melabathricum L. showed the presence of many phytochemical constituents which are responsible for antimicrobial property. Preliminary phytochemical profiling reflects essential information regarding the diversity of different classes of secondary metabolites such as alkaloids, flavonoids, steroids, saponins, tannins, reducing sugars etc. in the plant extracts. In the present study, qualitative tests for all three crude extracts showed significant indication about the presence of various secondary metabolites. Among the three selected samples, aqueous extract from leaves of Lawsonia inermis L. has shown the maximum antibacterial and antifungal activities against all tested microorganism than aqueous extract from leaves of Oroxylum indium (L.)Vent and Melastoma melabathricum L. From the result data, aqueous extract from leaves of Oroxylum indium (L.)Vent showed more zone of inhibition towards gram positive bacterial strain than gram negative bacteria and fungi. The aqueous extract from leaves of Melastoma melabathricum L. showed more zone of inhibition towards Bacillus subtilis and Escherichia coli bacteria strain. Therefore, it suggests that the aqueous extract from leaves of Lawsonia inermis L. Oroxylum indium (L.)Vent and Melastoma melabathricum L. can be a source of traditional medicine to be used i

REFERENCES

- Akinpelu, DA.,and TM. Onakoya 2006. "Antimicrobial Activities of Medicinal Plants used in Folklore Remedies in South-Western. Afri". J.Biotechnol. 5:1078-1081.
- Ayinde BA., EK.Omogbai, FC.Amaechina. 2007. " Pharmacognosy and Hypotensive Evaluation of Ficus exasperata Vahl (Moraceae) leaf". Acta Pol Pharm 64: 543-546.
- Bailey, L.H., Bailey, E.Z. 1976. Hortus Third: A concise dictionary of plants cultivated in the United States and Canada. New York: Macmillan. ISBN 978-0025054707
- Bhattacharje, SK. 2005. "Use of Flavours and Fragrances". In: Bhattacharje SK, editor. Handbook of aromatic plants. 2nd ed. Jaipur: Pointer Publishers;
- Burkill, I. H. 1996. "A Dictionary of Economic Products of Malay Peninsular." Ministry of Agriculture and Cooperatives, Kuala Lumpur, Malaysia
- Cuong NX, PT. Binh, MTT. Thuy,NH. Nami, HLT. Anh, NT. Dati ,NP.Thao, PV. LM. Kiem, Huong, 2009. "Isolation of Lawsonia inermis Leaves and Synthesis of its dimer Derivative by cyclic voltammetry". J Chem, 47:228–232
- Fazlin, A. S. M., Z. Ahmad, and H. H. Lim. 2002. "Compendium of Medicinal Plants Used in Malaysia". 2, Herbal Medicine Research Centre, Institute for Medical Research, Kuala Lumpur, Malaysia

- Jaganath I. B. and L. T. Ng, 2000. "Herbs: The Green Pharmacy of Malaysia." Vinpress Sdn. Bhd., Kuala Lumpur, Malaysia
- Kala,S., M. Johnson, N. Janakiraman, A. Anto Arockiaraj, S. Iyan Raj and Dorin Bosco, 2011. "Pharmacognostic and Phytochemical Studies on some selected ethnomedicinal plants of Tamilnadu, South India". Int. J. Med. Arom. Plants, 1, 2, , 89-94.
- Kenner D. and Requena, Y. 1996. "Botanical Medicine: A European professional perspective". Massachusetts. Paradign Publications. London.
- Khare CP. 2004. "Indian Herbal Remedies: Rational Western Therapy, Ayurvedic and other traditional usage, Botany". 4th. New York: Springer-Verlag Berlin Heidelberg; 340– 341.
- Koay, S. S. 2008. "Establishment of Cell Suspension Culture of Melastoma malabathricum L. for the Production of Anthocyanin." Ph.D. thesis, University Sains Malaysia, Pulau Pinang, Malaysia
- Latiff, A. and A. H. Zakri, 2000. "Protection of Traditional Knowledge, Innovations and Practices: The Malaysian Experience in the UNCTAD Expert Meeting on Systems and National Experiences for Protecting Traditional Knowledge, Innovations and Practices." Geneva, Switzerland
- Leal LKAM., AAG .Ferreira, GA. Bezerra, FJA. Matos, GSB. Viana, 2000. "Anticonceptive, Anti-inflammatory and Bronchodilator Activities of Brazilian medicinal plants containing coumarin: a comparative study". Journal of Ethnopharmacology.,70,(2):151-9. DOI:10.1016/S0378-8741(99)00165-8.
- Ling, K. H., C. T. Kian, and T. C. Hoon, 2009. "A Guide to Medicinal Plants". An Illustrated, Scientific and Medicinal Approach, World Scientific, Singapore
- Malinow, MR., P.McLaughlin, GO.Kohler, and AL. Livingstone, 1977. "Alfalfa saponins: a Family of substances potentially useful for treatment of hypercholesterolemia". J. Clin. Res. 25: 974-979.
- Okwu DE. and Josiah, C. 2006. "Evaluation of the Chemical Composition of Two Nigerian Medicinal Plants". *Afri. J.Biotechnol.* 5 (4):357-361
- Okwu DE., ME.Okwu, 2004. "Chemical Composition of Spondia smombin Linn. Plants parts". J Sust Agric Environ 6: 140-147.
- Olaleye, MT. 2007. "Cytotoxicity and Antibacterial activity of Methanolic extract of Hibiscus sabdariffa ". J. Med. Plants Res. 1(1):009-013.
- Raffauf, RF. 1996. "Alkaloids: A Guide to Their Discovery and Distribution". Hawkworth Press, Inc. New York.
- Rajenderan, M. T. 2010. "Ethno medicinal uses and antimicrobial properties of Melastoma malabathricum," SEGi Review, 3, 34–44
- Rubiay, KK, Al., Jaber NN, Al-Mhaawe, BH, Alrubaiy, LK. 2008. "Antimicrobial Efficiency of Henna Extract". Oman Med J ,23.253–256
- Sajem, A. L. and K. Gosai, 2006. "Traditional use of Medicinal plants by the Jaintia tribes in North Cachar Hills district of Assam, northeast India." Journal of Ethnobiology and Ethnomedicine, 2, 33
- Strasser, P.K., S. Anniyev, T and J. Greeley, K. More, C. Yu. 2010. "lattice strain control of the activity in dealloyed core-shell fuel cell catalyst." Nat Chem 2. 454-601
- Syamsudin I, H.Winarno 2008. "The Effect of Inai (Lawsonia inermis) Leaves Extract on Blood Sugar Level: An Experimental Study". *Res J Pharmacol*, 2:20–23.

- Vogel, A.I.1978. Vogels Text Book of Practical Organic Chemistry, 4th Edition, 1984 Reprint LRS/Londons, England, 280
- Warrier PK, VP. Nambiar, C. Ramankutty 1995. "Indian Medicinal Plants: A Compendium of 500 species". Chennai: Orient Longmann Pvt. Ltd,3:303.
- Warrier PK, VP. Nambiar, C.Ramankutty, R.Vasudevan 1995. "Indian Medicinal Plants: A Compendium of 500 species". 1st,Chennai: Orient Longmam Private Ltd;. Oroxylum indicum; 186–190.
- Wong, W. 2008. "Melastoma malabathricum: Too Beautiful to Be Called a Weed, Green Culture". Singapore,

Yoganarasimhan SN. 1996. "Medicinal Plants of India". Karnataka; Bangalore ,Interline Publishing;. 1, 366–367

- Yoganarasimhan SN. 1996. "Medicinal Plants of India". Karnataka; Bangalore, Interline Publishing, 1, 366–367
- Zakaria, Z. A., M. N. R. N. S. Raden, G. Hanan Kumar, Z.D.F. Abdul Ghani, M.R. Sulaiman, G.Rathna Devi, A.M. Mat Jais, M.N. Somchit, C.A. Fatimah, 2006. "Antinociceptive, Anti-inflammatory and Antipyretic Properties of Melastoma malabathricum Leaves Aqueous Extract in Experimental Animals". *Canadian Journal of Physiology* and Pharmacology, 84, 12, 1291–1299.
