

Available online at http://www.journalcra.com

INTERNATIONAL JOURNAL OF CURRENT RESEARCH

International Journal of Current Research Vol. 12, Issue, 02, pp.10303-10308, February, 2020

DOI: https://doi.org/10.24941/ijcr.38164.02.2020

RESEARCH ARTICLE

LEAF ANATOMICAL STUDIES OF ATALANTIA RACEMOSA WIGHT EX HOOK., AN IMPORTANT MEDICINAL PLANT OF TIRUVANNAMALAI, TAMIL NADU, INDIA

*Parthipan, G. and Dr. Shanthi, K.

PG and Research Department of Botany, Government Arts College, Tiruvannamalai-606603

ARTICLE INFO	ABSTRACT
Article History: Received 24 th November, 2019 Received in revised form 10 th December, 2019 Accepted 09 th January, 2020 Published online 28 th February, 2020	Atlantia racemosa (Rutaceae) is a small tree or shrub, differing from the other species of Atlantia and it is always unarmed, found in the lower mountain regions of Ceylon, in Southern India and in the western peninsula from the Concan to Travancore. Plant fruit pulp paste works as an antidote to Cobra bite while the leaf juice is taken internally to treat acidity. Leaf decoction is externally applied on itching skin. Seed oil of this plant is used to cure paralysis and chronic rheumatism. Anatomical characters are potentially useful to identify small fragments of commercially important herbal products as well as toxic plants that may be of relevance in forensic investigations. In this investigation, we screen the taxonomical characters execute the identification of <i>Atalantia racemosa</i> with anatomical studies of leaf in order to provide a scientific basis for this important medicinal plant
Key Words:	
Anatomical, Adulteration.	which these observations on microscopical characters confirmed that <i>A. racemosa</i> has apparent anatomical distinctiveness and peculiarities which differentiate with any adulterant taxa.
Conversion to 2020 Danthings and Chant	his This is an energy and a distributed under the Countries Commons Attribution License which memory internet internet

Copyright © 2020, Parthipan and Shanthi. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Citation: Parthipan, G. and Dr. Shanthi, K. 2020. "Leaf anatomical studies of Atalantia racemosa Wight ex Hook., an important medicinal plant of Tiruvannamalai, Tamil nadu, India.", International Journal of Current Research, 12, (02), 10303-10308.

INTRODUCTION

Plant anatomy or phytotomy is the general term for the study of the internal structure of plants, which provides characters such as trichomes, stomata, cuticular pattern, leaf venation, wood anatomy, growth rings etc. to aid in species identification and in performing physical matches of evidence. An increasing number of indigenous medicinal plants are being used both in the formal and informal economy so that the positive identification of species and infraspecific taxa has become important. Unfortunately very few of these plants have been studied anatomically. Plants are often the cause of fatalities due to the inappropriate use of herbal medicines or the use of misidentified plants (or sometimes intentionally used in homicide and suicide). Anatomical characters can be useful as supportive evidence in forensic work to help identify the plant species in question/anatomical evidence may often give additional clues as to the identity of the plant when medical diagnosis/ chemical forensic studies turn out to be inconclusive. Anatomical characters of potential diagnostic value will be presented for a selection of herbal medicine. Atlantia racemosa is a small tree or shrub, differing from the other species of Atlantia in that it is always unarmed. It is found in the lower mountain regions of Ceylon, in Southern India and in the western peninsula from the concan to Travancore1-3.

*Corresponding author: Parthipan, G.,

PG and Research Department of Botany, Government Arts College, Tiruvannamalai-606603.

In February the plant produces its white flowers which are arranged in short but distinct racemes with the peduncles a little less than a quarte of an inch in length. The berry is globular- ovoid, three- fourths of an inch wide with a long epiculus, four-celled, four seeded". (Trimen, Handbook of the flora of Ceylon and Hooker Flora of British India) introduced for the Citrus breeding work of the office of crop Physiology and Breeding Investigations for distribution later Jhon De Britto and Mahes, 2017; Ranade and Nair, 2009). Atlantia racemosa (Rutaceae) plants material collected from Kannalam, Villupuram District, Tamil nadu, India. Habit : Shrubs or small trees ca. 4m tall. Trunk\bark : Bark brownish, smooth; blaze yellowish. Branchlets : Branches with long spines; Branchlets terete, glabrous. Leaves : Leaves simple, alternate, spiral; petiole 0.5-1 cm long, planoconvex in cross section, glabrous, articulate; lamina 4.5-9 x 2.5-5 cm, elliptic to elliptic-ovate, apex emarginate, base acute to rounded, margin entire, coriaceous, glabrous, glandular punctate, drying olive green; midrib raised above; secondary nerves 10-18 pairs; tertiary nerves medially ramified. Flowers: Inflorescence axillary short cymes; flowers white; pedicel 0.4 cm long; stamens connate. Fruit& seed: Berry globose, to 1.9 cm long; seeds 4. Ecology: Undergrowth in evergreen forests up to 1000 m. Distribution: Peninsular India and Sri Lanka; in the Western Ghats- South, Central and south Maharashtra Sahyadris. Oral administration of Atlantia racemosa fruit pulp paste works as an antidote to Cobra bite (Sekar et al., 2011). The leaf juice is taken internally to treat acidity (Devi Prasad et al., 2013).

Leaf dodoction is externally applied on itching skin . oil from fruits is used to cure paralysis and chronic rheumatism (Sukumaran and Raj, 2010 & 2017). The leaves contain rich antioxidant molecules on natural basis. Also new antimicrobial drug can be developed for treating various disease from the selected plants (Saraswathy et al., 2017). Fruit pickles by tribes (Kudavul and Dixi, 2009). Antiviral activity has been observed in Ranikheit disease virus in the crude extracts of the plant excluding the root (Bhakuni et al., 1971). Leaf, rhizome and seeds are used inflammation diarrhea paralysis and chronic rheumatism arome therapy (Jayapal et al., 2014). Leaf, bark and fruit are cure digestive and cough (Symydurai et al., 2012). The leaf essential oil of Atlantia racemosa shows maximum (Larvicidal agents) activity against the three selected mosquito species namely Culex quinque fasciatus, Anapheles stephenst and Aedes aegypti (Arun et al., 2015).

MATERIALS AND METHODS

ANATOMICAL STUDIES

Collection of plant materials: Leaves of Atlantia racemosa (Rutaceae) for the proposed study were collected from Kannalam, Villupuram District, Tamil nadu, India and care was taken to select healthy plants and for normal organs. The identity of the plant specimens was confirmed by the use of local Floras (Gamble, 1957; Henry et al., 1987). The botanical identify was authenticated by Botanical Survey of India (BSI), Coimbatore, Tamil anadu, India.. Herbarium specimens of this plant (Fig.1) was deposited at PG and Research Department of Botany, Government Arts College, Tiruvannamalai, Tamil nadu, India. The required samples of different organs were cut and removed from the plant and fixed in FAA (Formalin - 5 ml + Acetic acid - 5 ml + 70 % Ethyl Alcohol - 90 ml) as per the schedule given by Sass (1940). Infiltration of the specimens was carried by gradual addition of paraffin wax (melting point 58-60°C) Tertiary Butyl Alcohol (TBA) until TBA solution attained super saturation. The specimens were cast into paraffin blocks.

Sectioning: The paraffin embedded specimens were sectioned with the help of Rotary Microtome. The thickness of the sections was 10-12 µm. De-waxing of the sections was done by customary procedure (Johansen, 1940). The sections were stained with Toluidine blue as per method published by O'Brian et al (1964). Since Toluidine blue is a polychromatic stain, the staining results were remarkably good and some cytochemical reactions were also obtained. The dye rendered pink colour to the cellulose walls, blue to the lignified cells, dark green to suberin, violet to the mucilage, blue to the protein bodies etc. Wherever, necessary sections were also stained with safranin and Fast-green and IKI (for starch). For studying stomatal morphology and venation pattern paradermal sections (sections taken parallel to the surface of leaf) as well as clearing of the leaf with 5 % sodium hydroxide or epidermal peeling by partial maceration employing Jeffrey's maceration fluid (Sass, 1940) were employed. Glycerine mounted temporary preparations were made for macerated materials. Powdered materials of different parts were cleared with NaoH and mounted in glycerine medium after staining. Different cell component were studied and measured.

Photomicrographs: Microscopic descriptions of tissues are supplemented with micrographs wherever necessary.

Photographs of different magnifications were taken with Nikon Labphot 2 microscopic unit. For normal observations bright field was used. For the study of crystals, starch grains and lignified cells, polarized light was used as they appear bright against dark background. Magnifications of the figures are indicated by the scale bars. Descriptive terms of the anatomical features are as given in the standard anatomy books (Esau, 1960, 1964).

RESULTS AND DISCUSSION

According to Metcalf and Chalk (1957) microscopical characters are necessary to establish the botanical identity of commercial samples of medicinal plants and play an important role in checking adulteration and substitution. Anatomical and Histochemical studies of leaves of

Atlantia racemosa: The leaf is bifacial comprising distinct adaxial and abaxial sides. In sectional view, the lamina is smooth and even. The midrib in thick and wide assuming semicircular adaxial past and slightly convex abaxial part (Fig.1.1). The midrib is 600µm thick. The adaxial dome shaped part is 550µm wide. The epidermal layer along the adaxial part consists of vertically oblong small cells with thick cuticle. The abaxial epidermal cells are small less prominent, thick called and darkly stained (Fig.1.2). The ground tissue of midrib consists of small, polyhedral the thick compact parenchyma cells. The vascular system of the midrib double starved, these an adaxial slightly curved thick plate vascular strand and an abaxial wide arch of vascular strand. Both adaxial and abaxial vascular strands are collateral. The phloem units are located on the outer (adaxial and abaxial) ends of the xylem. The xylem segments comprise several long, parallel, vertical rows of thick walled lignified angular xylem elements. The protoxylem cells are towards the center (Fig.3.3). The phloem strands are thick and dense and it includes the strands of compact parallel lines of angular cells. Thick and wide segments sclerenchyma cells are abutting closely the phloem strands. The vascular system, excluding sclerenchyma layers, measures 300µm both along the vertical plane and horizontal plane wide, circular lysigenous secretary cavities common the ground tissue. The cavities be 130µm wide.

Lamina: Lamina is smooth and even on to the adaxial and abaxial surfaces. The lamina is 250μ m thick. The adaxial epidermis includes thick squarish cells with thick cuticle. The epidermis is 20μ m thick (Fig 2.1). The abaxial epidermal cells are small, thick walled with their cuticle. The mesophyll tissue is differentiated into adaxial band of two layers of pillar like thin and compact palisade cells and tick abaxial zone of about eight layers of irregular and circular loosely arranged spongy parenchyma cells. Lateral vein are often seen in the lamina. The vein are more or less vertically elliptical and are composed of small xylem and phloem stands caped by very thick upper and lower selerenchyma masses.

Leaf margin (Fig. 2.2): The marginal part of the leaf is thick and conical. It is 150μ m thick. The structure of the leaf margin is similar to the mid part of the lamina. There is a thick double strands of palisade cells and thick compact spongy mesophyll layers. Calcium oxalate druses are abundant in the mesophyll tissues of the leaf. The druses are solidar; they are diffuse in distribution. The cell bearing the druses are not modified in size and shape (Fig. 2.3) The druses are 20μ m thick.



Fig.1.1. T.S of leaf through midrib



Fig.1.2. T.S of leaf midrib enlarged



Fig.1.3. Vasculpar bundle of the midrib (Ab ph-Abaxial phloem; Abs- Abaxial side; Abx- Abaxial phloem; Adph-Adaxial phloem; Ads-Adaxial side; Adx-Adaxial xylem; Cu-cuticle; Ep-Epidermis; La-Lamina; Mr-Midrib; Sc-Secretory cavity; VB-Vascular bundle)

Epidermal Tissues: The epidermal cells and the stomata were studies from paradermal sections. The cells appear in surface view. The adaxial epidermis consists of polygonal cells with highly thick walls. The anticlinal walls are straight. The cells have abundant simple circular pits. (Fig 3.1, 2,3). The pits are diffuse in distribution. Due to the presence of pits, the anticlinal walls appear beaded (Fig 3.2). The adaxial epidermis is apostomatic (without stomata). The abaxial epidermal cells are also polygonal in outside. The cells are small; they have thick and straight anticlinal walls. The cells are densely pitted; the pits are simple and circular (Fig 4.1, 2, 3). The abaxial epidermis is μ stomatiferous. The stoma are diffuse in distribution (Fig 4.1,2). The guard cells circular in surface view with semicircular guard cells.



Fig.2.1. T.S of lamina



Fig.2.2. T.S of lamina – marginal part



Fig.2.3. Crystal distribution in the mesophyll (AbE-Abaxial Epidermis; AdE-Adaxial Epidermis; Cr-Crytals; Ep-Epidermis; LM-Leaf Margin; PM-Palisade Mesophyll; SM-Spongy Mesophyll; VB-Vascular Bundle; Sc-Sclerenchyma)

The stomata are cyclocytic type. Each stomata is surrounded by a circle of four or five subsidiary cells. The subsidiary cells are rectangular and curved (Fig 4.1,3; 5.2). The stomata are 30 X 30 μ m in size.

Venation pattern (Fig 6.1,2) The venation of the lamina is densely reticulate. The primary lateral veins are very thick; the secondary vein and the vein-lets are also fairly thick. The vein-islet are elongated and narrow. They are bordered by thick vein boundaries (Fig 6.1). The vein terminations are many in each islet. The terminations are either unbranched or branched once or twice (Fig 6.2). The terminations are slightly curved.

Starch grains (Fig 7.1,2) Starch grains are abundantly in the mesophyll cells of the leaf. The starch grain simple type with central hilum and + shaped polarimark.



Fig.3.1. Paradermal sections of the adaxial epidermis



Fig. 3.2. Epidermal cells in surface view



Fig. 3.3. Showing thick beaded anticlinal walls and simple pits. (AW-Anticlinal walls; EC-Epidermal Cells; Pi-Pits)



Fig. 4.1. Abaxial epidermis in surface view



Fig. 4.2. Leaf paradermal setion



Fig. 4.3. Leaf paradermal setion (AW-Anticlinal walls; EC-Epidermal cells; GC-Guard cells; SC-subsidiary cells; St-Stomata)



Fig. 5.1& 2. Stomata enlarged showing cyclocytic subsidiary cells (AW-Anticlinal walls;; Gc-Guard cells; Pi-Pits; Sc-Subsidary cells)



Fig.6.1 Venation pattern of the lamina



Fig 6.2. Vein-islet and vein terminatious enlarged (VI-Venation Islet; VT- Vein Terminatious)



Fig.7.1. Starch grains in the seaf mesophyll (as seen under polarized light) 3Foliar scleroid in the leaf.

The starch grains are $10\mu m$ in diameter. Foliar selereids (Fig. 7.3) Sclerides are often seen in the leaf. They are commonly seen associated with the veins. The scelerids are long, thin, unbranched curved thread like and lignified.

Petiole (Fig 8.1, 2,3). The petiole is semicircular in sectional view. The adaxial side is flat and the abaxial side in convex (Fig8.1). The petiole is 1.3mm in horizontal plance and 1.1mm in vertical plane. The epidermal small epidermal cells with smooth the thick cuticle. The ground tissue of the petiole is homogenous, parenchymatous and thin walled. These are wide, circular secretary cavities distributed in the ground tissue. The cavities 40μ m wide (Fig 8.2). There is a single vascular bundle comprising two segments (Fig 8.3). On the adaxial part is a flat thick vascular segment which consists of short radial rows of xylem and xylem fibres and adaxially located phloem unit.



Fig. 8.1. T.S. of petiole - entire view



Fig. 8.2. T.S. of petiole – A sector



Fig. 8.3. T.S. of petiole – vascular strand enlarged (Ab Ph- Abaxial Phloem; Abx- Abaxial xylem; AdPh-Adaxial phloem; Adx-Adaxial xylem; Ep-Epidermis, Ph-Phloem; SC-Secretary Cavity; X-Xylem)

The abaxial segment includes a thick are shaped xylem strand and wide and thick are of phloem formed on the lower part of the xylem (Fig 8.3) The entire vascular completed in enclosed within a thin discontinuous layer of sclerenchyma cells. The vascular structures measure $480\mu m$ in horizontal plane and $350\mu m$ in vertical plane.

Summary and Conclusion

Atalantia racemosa a rare species in Tailnadu is known for its ethnomedicobotanical relevance. Few studies have been carried out by other researchers which have fragmentary information on specific parts. No Morphological and antatomical studies have been done till date. Our investigation highlights the botanical standardization of whole plant of *Atalantia racemosa*, which is claimed as potent ethnodemicinal plant in the form of drug. This work presents a descriptive study of the anatomical characters of leaf.

We conclude that, the anatomical findings *viz.*, *xy*lem and phloem fibers, vessel elements, epidermal tissues, calcium oxalate crystals, trichomes, and unique foliar architecture with micrographic information which has provided taxonomic value for identifying and classifying the desired plant taxon to the other against adulteration.

REFERENCES

Arun Das K., Suresh kumar J., Swamy PS. 2015. Larvicidal activity and leaf essential oil composition of three species of genus Atlantia from south India. *International Journal of Mosquito Research*, 2(3); 25-29.

- Bhakuni, D.S. et al., Indian Journal of Experimental Biology, 1971, 993.
- Devi Prasad AG., Shyma TB., Raghavendra MP. 2013. Plants used by the tribes for the treatment of digestive system disorders in wayanad district, Kerala. *Journal of Applied pharmaceutical science*, (3): 171-175.
- Esau K. 1960. *Anatomy of seed plants*. John Wiley and Sons, New York, U.S.A. 364.
- Esau K. *Plant Anatomy* (2nd edn.). Wiley Eastern Limited, New Delhi, India. 1964: 719.
- Gamble, J.S. 1921. "*Flora of Presidency of Madras*". Botanical Survey of India, Calcutta, II: 1169 and 1179.
- Henry, A.N., Kumari, G.R. and Chitra, V. 1987. "Flora of Tamilnadu", India Series I: Analysis. Botanical Survey of India, Southern circle, Coimbatore, II: 190-92.
- Jayapal J, Tangaelou AC., Panneerselvam A. 2014. Studies on the plant diversity of Muniandavar sacred groves of Thiruvaiyaru, Thanajavur, Tamil nadu, India. *Journal for drug and Medicines*, 6(1): 48-62.
- Jayapal J. Tangaelou AC, Panneerselvam A. 2014. Studies on the plant diversity of Muniandavar sacred groves of Thiruvaiyaru, Thanajavur, Tamil nadu, india. *Journal for drug and Medicines*, 6(1): 48-62.
- Jhon De Britto A, Mahes R. 2017. Exploration of kani tribal botanical knowledge in Agasthiyamalai Biosphere reserve. South India. *Ethanobotanical leaflet*, 11:258-265.
- Johansen DA. 1940. *Plant Microtechnique*. McGraw' Hill Book Co. Inc., New York. 183-195.
- Kadavul K., Dixit A.D. 2009. Ethanomedicinal studies of the woody species of Kalrayn and Shervarayan hills, Eastern Ghats, Tamil nadu. *Indian Journal of Traditional Knowledge*, 8(4): 592-597.
- O'Brien TP., Feder N., McGill ME. 1964. Polychromatic staining of plant cell wall by Toluidiner blue-O. *Protoplasma*, 59: 364-373.
- Ranade SA, Nair KN, Srivastava AP. 2009. Pushpagandan. Analysis of diversity amongst widely distribution and endemic Atlantia (Family Rutaceae) species form Western Ghats of India. *Physiology Mol. Biol. Plants.*, 135: 211-223.
- Samydurai P., Jagathesh kumar S., Aravinthan V., Thangapandian V. 2012. Survey of wild aromatic ethanomedicinal plants of Velliangiri hills in southern western ghats of Tamil nadu, India. *Journal of Medicinal and Aromatic Plants.*, 2(2): 229-234.
- Saraswathi K., Mahalakshmi B., Rajesh V., Arumugam P. 2017. In vitro evaluation of antioxidant and antimicrobial potential of leves of Atlantia racemosa wight ex Hook. *International Journals of Pharma Research and Health Science*, 5(6): 2031-37.
- Sass JC. 1958. Botanical Microtechnique, Oxford TBN Publishing Co., New Delhi. Print.
- Sekhar J., Penchala Pratap G. el al., 2011. Ethnic information on treatments for snake Bites in Kadapa district of Andhra Pradesh, Life sciences Leaflets (12): 368 – 375.
- Sukumaran S, Raj ADS. 2007. Rare endemic threatened trees and lians in the sacred groves of Kanyakumari district, India. 133(9):1254-1266.
- Sukumaran S, Raj ADS. 2010. Medicinal plants of sacred grooves in Kanyakumari district, southern western Ghat. *Indian Journals of Traditional Knowledge*, 9(2): 294-299.