



International Journal of Current Research Vol. 6, Issue, 11, pp.10077-10080, November, 2014

## RESEARCH ARTICLE

# CHANGES INDUCED BY FOLIAR APPLICATION OF SALICYLATE ON ENZYME ACTIVITIES IN VIGNA MUNGO (L.) HEPPER

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# **ARTICLE INFO**

#### Article History:

Received 18<sup>th</sup> August, 2014 Received in revised form 27<sup>th</sup> September, 2014 Accepted 09<sup>th</sup> October, 2014 Published online 30<sup>th</sup> November, 2014

#### Key words:

Enzyme Activity,
Catalase,
Peroxidase,
Polyphenol Oxidase,
Salicylic Acid,
Superoxide Dismutase Vigna Mungo,

### **ABSTRACT**

Salicylic acid (SA) is a growth regulator that promotes growth of plants under stress and non stress conditions. The present study was conducted to determine the effect of foliar applications of salicylic acid (SA) on enzyme activity of *Vigna mungo* (L.) Hepper. SA was applied at five different concentrations (50ppm, 100ppm, 150ppm, 200ppm 250ppm) during the growth period of 15 days old *Vigna* seedling. Activities of antioxidant enzymes such as catalase (CAT), nitrate reductase activity (NRA), peroxidase (POD) polyphenoloxidase (PPO) and superoxide dismutase (SOD) were determined in the fresh leaves obtained from 15days old seedlings. Foliar applied SA caused a significant increase in leaf SOD and POD activity. However, leaf CAT activity was decreased with all SA treated plants compare to control plants.

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# **INTRODUCTION**

Salicylic acid (SA) is a ubiquitous phenolic compound occurring in plants in very low amounts and has been reported to regulate the physiological processes in plants such as nutrient uptake, stomatal closure, inhibition of ethylene biosynthesis, chlorophyll synthesis, protein synthesis, photosynthesis and transpiration (Khan et al., 2003, Raskin 1992, Shakirova et al., 2003). The expression of pathogenesis related protein genes has been regulated by SA, suggesting its key role as a signal molecule by providing resistance against pathogen attack (Raskin 1992). As an elicitor, SA regulates the PAL enzyme activity, which as a biosynthetic enzyme, catalyzes biosynthetic reactions for forming defensive compounds (Sgarbi et al., 2003; Solecka and Kacperska, 2003; Zhao et al., 2005) and SA regulates the protective enzymes such as SOD and POD, which together increase a plant's tolerance to environmental stresses (Mutlu et al., 2009; Shi and Zhu, 2008; Shim et al., 2003; Thulke and Conrath, 1998). Exogenous SA can regulate the activities of intracellular antioxidant enzymes such as SOD, POX and increases plants tolerance to environmental stresses (Senaratna et al., 2000, Sakhabutdinova et al., 2004). Vigna mungo, also known as black gram or urdbean, is a leguminous crop grown in south East Asia.

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Centre for Research and Postgraduate Studies in Botany, Ayya Nadar Janaki Ammal College (Autonomous, College of Excellence), Sivakasi-626 124, Tamilnadu, India. It belongs to family fabaceae and subfamily faboideae. It originated in India and has been in cultivation since ancient times. The seeds contain about 20-25% of proteins and 40-47% of starch. It plays a very vital role in overcoming the protein caloric malnutrition especially in the developing country like India, where majority of population is vegetarian. It also has a superior mineral profile which makes it nutritionally more balanced. Moreover, its extracts have immuno stimulatory activity Solanki *et al.* (2010). The objectives of the present research was to improve our understanding of the effect of the various concentrations of SA applied to *Vigna mungo* plants as foliar spray on the catalase (CAT), nitrate reductase activity (NRA), peroxidase (POX) polyphenoloxidase (PPO) and superoxide dismutase (SOD) and its assimilation black in gram and to screen for the most effective concentration of SA.

# **MATERIAL AND METHODS**

#### Plant material

Healthy and uniform seeds of *Vigna mungo* (L.) Hepper variety were purchased from Agricultural Research Station, Kovilpatti and surface sterilized with 0.1% HgCl<sub>2</sub> for a minute and washed repeatedly in distilled water. Healthy seeds were selected and sown in pots containing mixture of red soil, black soil, and sand mixed in the ratio of 2:2:1. The seeds were allowed to germinate in dark for 48 h. The percentage of seed

germination was nearly 80%. Soon after emergence, the seedlings were shifted to daylight conditions. After (15 days) growth of seedlings, the seedlings were sprayed with different concentrations of SA (50ppm, 100ppm, 150ppm, 200ppm, 250ppm) using an atomic sprayer. The seedlings were sprayed with solutions until dropping. Each plant required about 10ml of spray solution. Salicylic acid (SA-2-hydroxybenzoic acid) was obtained from Sigma Chemical Co. (St. Louis, U.S.A), SA was initially dissolved in 100ul of dimethyl sulfoxide and concentrations of 5 x 10<sup>-6</sup>M to 100 x 10<sup>-6</sup>M (pH 6.5) were made up with distilled water containing 0.02% Tween-20 (Polyoxyethylene sorbitan monolaurate). Plants sprayed with 0.02% Tween-20 served as the control. The plants were arranged in a completely randomized design with three replicates (Khan et al., 2003).

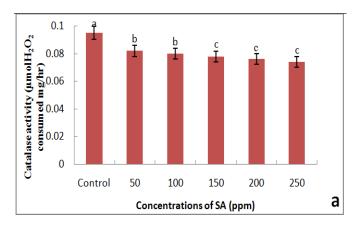
## Estimation of enzyme activity

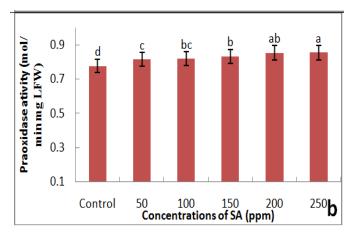
In vivo Nitrate Reductase (NR) activity was assayed according to Jaworski (1971) method Absorbance was measured at 540nm. The nitrate was estimated with the help of nitrite standard curve. To assay catalase activity, 3ml, phosphate buffer was added to 1ml of H<sub>2</sub>O<sub>2</sub> and 1ml of enzyme extract (Kar and Mishra, 1976). The reaction mixture was incubated at 25°C for 1 minute. The reaction was terminated by the addition of 1ml of H<sub>2</sub>So<sub>4</sub>. The reaction mixture was titrated against 0.01N KMN0<sub>4</sub>. The end point was the persistence of pink colour at least for 15 seconds. The catalase activity was expressed in micro moles H<sub>2</sub>O<sub>2</sub> catalyzed per unit time per mg protein. To assay peroxidase activity, the enzyme extract was added to pyrogallol which gets oxidized to coloured derivative in the presence of hydrogen peroxide (1% V). The amount of purpurogallin formed during the reaction was assayed spectrophotometrically Addy and Good man (1972). Polyphenol oxidase activity was analysed by colorimatric method (Mukherje et al., 1975). To 2ml of enzyme extract 3ml of 0.1m phosphate buffer (pH 6.0) was added and mixed thoroughly by inverting the cuvette and placed in calorimeter. Super oxide Dimutase (SOD) activity was analyzed by Bowler et al. (1992) method. The absorbance was measured at 560nm.

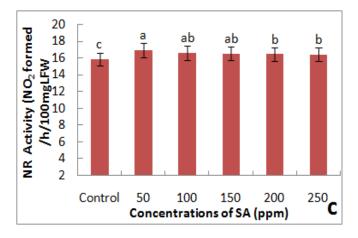
**Statistical analysis**: The experiments were performed in a randomized order. Datas were expressed as means of three replicates with standard error. Statistical assays were carried out by one-way ANOVA using Tukey's test to evaluate whether the means were significantly different, taking P < 0.05 as significant.

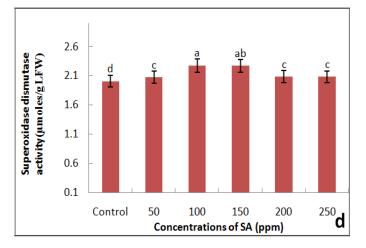
# **RESULTS AND DISCUSSION**

Catalase, one of the antioxidant scavenging enzyme analysed in the present study was found to be decreased with all concentration of salicylic acid treated at 15 days old *Vigna* seedlings compared to control (Fig1-a). The catalase activity was found to be increased in control plant. There is an evidence that SA will decrease the CAT activity (Dat *et al.*, 2000; Shi *et al.*, 2006; Shi and Zhu, 2008; Shim *et al.*, 2003), but in Kentucky bluegrass (He *et al.*, 2005) and in wheat (Agarwal *et al.*, 2005), SA actually increased the CAT activity. The greatest responses were obtained in plants sprayed with 200ppm and 250ppm of SA, with significant increases observed in POD (9% and 10%).









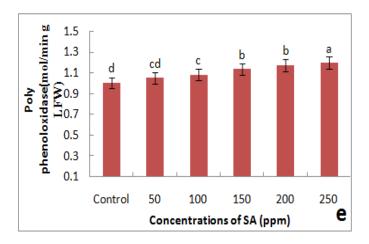


Fig. 1-(a,b,c,d,e,). Effect of Salicylate spray on enzyme activity in intact seedlings of *Vigna mungo* (L.) Hepper. The concentrations of SA were 50,100,150,200,250ppm. Each seedlings received 10ml / plant of SA spray given during the early hours of the day after 15 days old seedlings. The measurements were made 15 days old seedlings. Each value represent the mean of three independent measurements (Mean $\pm$ SE, n=3). Bars carrying different letters are significantly different at p < 0.05

Peroxidase activity was increased with different concentrations of SA treated 15 days old Vigna seedlings compared to control (Fig1-b). Both the catalase and peroxidase act as the scavenger of superoxidase radicals. The treatment of millet seeds with salicylic acid caused the increase of peroxidase activity at all stages of experiment. The increase of peroxidase activity under the influence of salicylic acid is shown earlier too with the example of other cereals - barley (Ananieva, Popova, 2002) and wheat (Kolupaev et al., 2010). Salicylic acid can also exert the various (both activating and inhibiting) influence on the activity of peroxidase (Ruffer et al., 1995; Ananieva, Popova, 2002; Kolupaev et al., 2010), Application of 50ppm (7%) SA was significantly superior to other concentrations of Vigna seedlings, although lower nitrate reductase activity was observed in control plants (Fig 1-c). Our results corroborate the findings of many other related studies. In particular, they are consistent with those of Fariduddin et al. (2003) found that low SA concentrations increased nitrite reductase activity in Brassica juncea, whereas higher concentrations were inhibitory. It can therefore be assumed that SA concentration plays an important role in regulating nitrate reductase activity, with lower concentrations enhancing nitrate reductase protein and higher concentrations decreasing it by affecting the above processes. Increases in nitrate concentration and in turn nitrate reductase activity due to exogenous SA treatment under normal growth conditions have been reported previously and strongly support our observed results Hayat et al. (2005). Nitrate reductase activity results was found to increase with decreased in SA concentrations (Fig1-c).

SOD of 15 days old seedlings was found to be increased with the concentration of hormone. The SOD of the seedlings increased to 100ppm (13%), 150ppm (12%), were observed (Fig 1-d). The SOD is maximum around 100 and 150ppm concentrations. After that results was compared to control respectively (Fig 1-d). SOD is a major scavenger of superoxide radicals and the enzymatic action results in the formation of

H<sub>2</sub>O<sub>2</sub> that is then converted to H<sub>2</sub>O and O<sub>2</sub> by POD and CAT (Alscher *et al.*, 2002; Takahashi and Asada, 1983). Salicylic acid can increase SOD and POD activities with the resulting increase of AOS scavenging to protect plants from being injured (Mutlu *et al.*, 2009; Shi and Zhu, 2008).

Regarding polyphenoloxidase, a consistent increase at 200ppm (17%), 250ppm (19%) of SA on 15 days old seedlings were observed (Fig 1-e). The polyphenoloxidase was maximum around 200, 250ppm concentrations. The minimum results were around 50 and 100ppm concentrations, the results were compared to the respective controls. (Fig. 1-e). The effects of SA on apoplastic enzyme activity in wheat plant leaves, was found to increase with SA treatment, while apoplastic catalase enzyme activity decreased, apoplastic peroxidase and polyphenol oxidase activities increased (Tasgin *et al.*, 2003).

# **REFERENCES**

Addy, S.K. and Good man, R.N. 1972. Polyphenoloxidase and peroxidase activitity in apple leaves inoculated with a virulent or an avirulent strain for *Erwinia amylovora*, Indian phytopatho 25:575-579.

Agarwal, S., Sairam, R.K., Srivastava, G.C., Tyagi, A. and Meena, R.C.2005. Role of ABA, Salicylic acid, Calcium and hydrogen peroxide on antioxidant enzymes induction in wheat seedlings. *Plant Sci.*, 169:559-570.

Alscher, R.G., Erturk, N. and Health, L.S. 2002. Role of superoxide dismutases (SODs) in controlling oxidative stress in plants. *J. Exp. Bot.*, 53, 1331–1341.

Ananieva, E.A. and Popova, L.P. 2002. Regulatory role of salicylic acid in paraqvat-induced oxidative damage in barley plants. Dokl. Bulg. AN, 55, 65-68.

Bowler, C., Van Montagu, M., Inzé, D. 1992. Superoxide dismutase and stress tolerance. *Ann. Rev. of Plant. Physiol. and Plant Mol. Biol.*, 43, 83–116.

Chen, J.Y., Wen, P.F., Kong, W.F., Pan, Q.H., Zhan, J.C., Li, J.M., Wan, S.B. and Huang, W.D. 2006. Effect of salicylic acid on phenylpropanoids and phenylalanine ammonialyase in harvested grape berries. *Postharvest Biol. Technol.*, 40, 64–72.

Dat, J.F., Lopez, D.H., Foyer, C.H. and Scott, I.M. 2000. Effects of salicylic acid on oxidative stress and thermo tolerance in tobacco. *J. Plant Physiol.*, 156, 659–665.

Fariduddin, Q., Hayat, S. and Ahmad, A. 2003. Salicylic acid influences net photosynthetic rate, carboxilation efficiency, nitrate reductase activity and seed yield in *Brassica juncea*. *Photosynthetica.*, 41: 281–284.

Hayat, S., Fariduddin, Q., Ali, B. and Ahmad, A. 2005. Effect of salicylic acid on growth and enzyme activities of wheat seedlings. *Acta Agron. Hung.* 53, 433–437.

He, Y., Liu, Y., Cao, W., Huai, M., Xu, B. and Huang, B. 2005. Effects of salicylic acid on heat tolerance associated with antioxidant metabolism in Kentucky bluegrass. Crop Sci., 45, 988–995.

Jaworski, E.G., 1971. Nitrate reductase assay intact plant tissues. Biochem, *Biophy, Res. Commun.*, 43:1274-1279.

Kar, M. and Mishra, D. 1976. Catalase, peroxidase and polyphenol-oxidase activities during rice leaf senescence, *Plant physiol.*, 57:315-319.

- Khan, W., Prithiviraj, B. and Smith, D.L. 2003. Photosynthetic responses of corn and soybean to foliar application of salicylates. *J. Plant. Physiol.*, 160: 485-492.
- Kolupaev, Yu.Ye., Karpets, Yu.V., Yastreb, T.O. and Musatenko, L.I. 2010. Participation of peroxidase and superoxide dismutase in the increase of reactive oxygen species production by wheat coleoptiles at the action of salicylic acid. Fiziol. Biokh. Kul't. Rast, 42, 210-217. (In Russ.).
- Mukherjee, P.K. and Ghosh, J.J. 1975. Phenoloxidase activity in reaction to resistance of rice to infection by Helmin thosporium oryzae. *Sci. Culture*, 41:433-435.
- Mutlu, S., Atici, Ö. and Nalbantoglu, B. 2009. Effects of salicylic acid and salinity on apoplastic antioxidant enzymes in two wheat cultivars differing in salt tolerance. *Biologia Plantarum*, vol. 53, no. 2, p. 334-338.
- Raskin, I. 1992. Role of salicylic acid in plants. *Ann. Rev. Plant Physiol. Mol. Biol.*, 43: 39-463.
- Ruffer, M., Steipe, B. and Zenk, M.H. 1995. Evidence against specific binding of salicylic acid to plant catalase. FEBS Lett. 377:175-180.
- Sakhabutdinova, A.R., Fatkhutdinova, D.R. and Shakirova, F.M. 2004. Effect of salicylic acid on the activity of antioxidant enzymes in wheat under conditions of salination. *Applied Bioche and Microbiol*. 40:501-505.
- Senaratna, T., Tuochell, D., Bunn, T. and Dixon, K. 2000. Acetylsalicylic acid (aspirin) and Salicylic acid induce multiple stress tolerance in bean and tomato plants. Plant Growth Regul., 30, p.157-161.
- Sgarbi, E., Fornasiero, R.B., Lins, A.P., Bonatti, P.M. 2003. Phenol metabolism is differentially affected by ozone in two cell lines from grape (*Vitis vinifera* L.) leaf. Plant Sci., 165, 951–957.
- Shakirova, F.M., Sakhabutdinova, A.R., Bezrukova, M.V., Fatkhutdinova, R.A. and Fatkhutdinova, D.R. 2003. Changes in the hormonal status of wheat seedlings induced by salicylic acid and salinity. Plant Sci. 164: 317-322.

- Shi, O., Bao, Z., Zhu, Z. and Ying, O. 2006. Effects of different treatments of salicylic acid on heat tolerance, chlorophyll fluorescence, and antioxidant enzyme activity in seedlings of *Cucumis sativa* L. Plant Growth Regul., 48, 127–135.
- Shi, Q.H., Zhu, Z.J. 2008. Effects of exogenous salicylic acid on manganese toxicity, element contents and antioxidative system in cucumber. Environ. Exp. Bot., 63, 317–326.
- Shim, I.S., Momose, Y., Yamamoto, A., Kim, D.W., Usui, K. 2003. Inhibition of catalase activity by oxidative stress and its relationship to salicylic acid accumulation in plants. *Plant Growth Regul.*, 39, 285–292.
- Solanki, Y.B. and Jain S.M. 2010. Immunostimulatory effect of *V. mungo* L. extract in male Sprague- Dawley rats. *J. Immunotoxicol.*, 7 (3): 213-218.
- Solecka, D., Kacperska, A. 2003. Phenylpropanoid deficiency affects the course of plant acclimation to cold, *Physiol. Plant.*, 119: 253-262.
- Takahashi, M.A., Asada, K. 1983. Superoxide anion permeability of phospholipids membranes and chloroplast thylakoids. *Arch. Biochem. Biophys.*, 226, 558–566.
- Tasgin, E., Atici, O., Nalbantoglu, B. 2003 Effects of salicylic acid and cold on freezing tolerance in winter wheat leaves. *Plant Growth Regul.*, 41, p. 231-236.
- Thulke, O., and Conrath, U. 1998 Salicylic acid has a dual role in the activation of defence-related genes in parsley. *Plant J.*, 14(1): 35–42.
- Zhao, J., Davis, L.T., Verpoort, R. 2005. Elicitor signal transduction leading to production of plant secondary metabolites. *Biotechnol. Adv.*, 23, 283–333.

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